

## Development and validation of a new and economical stability indicating RP-HPLC method for cefixime trihydrate

Ceema Mathew<sup>1,2,\*</sup>, Bellamkonda Swathi<sup>1</sup>, Makula Ajitha<sup>2</sup>, Puvvadi Sathesh Babu<sup>1</sup>

<sup>1</sup>Gokaraju Rangaraju College of Pharmacy, Osmania University, Bachupally, India, <sup>2</sup>Jawaharlal Nehru Technological University, Hyderabad, India

The present work describes the development of a new high performance liquid chromatographic (HPLC) method for the determination of Cefixime trihydrate under different stress conditions as specified by ICH. For the analysis, a Phenomenex (250 x 4.6 mm, 5 µm particle size) ODS column and a SPD 20 A UV detector at 289 nm was used. The selected mobile phase was 10 mM disodium hydrogen phosphate (with 0.5% TEA, pH adjusted to 6.3 with OPA) and methanol in the ratio of 75:25 (v/v) in isocratic mode at a flow rate of 1 mL.min<sup>-1</sup>. The linear regression analysis data for the calibration plots showed good linear relationship with  $r^2 = 0.9997$  in the concentration range of 5-100 µg.mL<sup>-1</sup>. The stress degradation was performed using acid, alkali, water, hydrogen peroxide and uv light.

**Uniterms:** High Performance Liquid Chromatography/quantitative analysis. Degradation. Stability-indicating. Cefixime trihydrate/determination.

O presente trabalho descreve o desenvolvimento de um novo alta performance cromatografia líquida (HPLC) método para a determinação de cefixima tri-estresse sob diferentes condições, conforme especificado pelo ICH. Para a análise, a Phenomenex (250 x 4,6 mm, 5 µm de granulometria) ODS coluna e a SPD 20 um detector de UV em 289 nm foi utilizado. A fase móvel selecionado foi de 10 mM hidrogenofosfato dissódico (com 0,5% TEA, o pH ajustado para 6,3 com OPA) e de metanol em razão de 75:25 (v/v) no modo isocrático com uma taxa de fluxo de 1 mL.min<sup>-1</sup>. A análise de regressão linear para dados da calibração parcelas apresentaram boa relação linear com  $r^2 = 0,9997$  no intervalo de concentração de cerca de 5 100 µg.mL<sup>-1</sup>. Degradação do estresse foi realizado utilizando um ácido, alcalino, a água, o peróxido de hidrogênio e luz uv.

**Unitermos:** Cromatografia líquida de alto desempenho/análise quantitativa. Degradação. Indicador de estabilidade. Cefixima triidratada/determinação.

### INTRODUCTION

Cefixime trihydrate (Chemical structure is given in Figure 1) [(6R,7R)-7-(2-(2-amino-4-thiazolyl) glyoxylamido)-8-oxo-3-vinyl-5-thia-1-azabicyclo[4.2.0] oct-2-ene-2-carboxylic acid, 72-(Z)-[O-(carboxymethyl) oxime] is an orally absorbed third generation cephalosporin antibiotic that was approved by the U.S. Food and Drug Administration in 1997 for the treatment of mild to moderate bacterial infections. It has a broad antibacterial spectrum against various Gram-positive and Gram-negative bacteria, including Haemophilus influenzae,

Neisseria gonorrhoeae, Escherichia coli and Klebsiella pneumoniae resistant to ampicillin, cephalexin, cefaclor and trimethoprim - sulfamethoxazole. It is used for the treatment of susceptible infections, including gonorrhea, otitis media, pharyngitis, lower respiratory-tract infections such as bronchitis and urinary-tract infections (Brogden, Richards, 1989; Sweetman, 2002; Zahra Talebpour *et al.*, 2013).

Cefixime degrades under storage conditions (temperature and relative humidity) and its degradation products can cause undesirable side effects in patients (Kitamura *et al.*, 1989; Kitamura *et al.*, 1990). A detailed literature survey has shown that cefixime has been studied by various analytical methods like spectrophotometric (El-Walily *et al.*, 2002; Attimarad *et al.*, 2011), fluorimetric

\*Correspondence: Ceema Mathew. Gokaraju Rangaraju College of Pharmacy. Bachupally, Hyderabad-90, India. E-mail: nirujose@gmail.com

(Bebawy, El-Kelani, Fattah, 2003), voltammetric (Golcu, Dogan, Ozkan, 2005; Jain *et al.*, 2010), HPLC (Dhoka, Sandage, Dumbre, 2010; Arshad *et al.*, 2009; Khan *et al.*, 2008; Shah, Pundarikakshudu, 2006; Manna, Valvo, 2004; Meng *et al.*, 2005; Gonzalez-Hernandez *et al.*, 2001; Khandagle, 2011), and HPTLC (Singh, Maheshwari, 2010; Eric-Jovanovic *et al.*, 1998; Pawar *et al.*, 2010).

Although different analytical methods are available, a more economical stability indicating analytical method is developed for cefixime. We have forcefully degraded the drug (standard) under different stress conditions and developed an HPLC method that can differentiate the pure drug from its degradants. In an attempt to practice green chemistry and also to develop a cost effective analytical method, we have tried to minimise the use of organic solvent (ie methanol to 25%).

The International Conference on Harmonization (ICH) guideline entitled “Stability Testing of New Drug Substances and Products” requires that stress testing be carried out to elucidate the inherent stability characteristics of the active substance (ICH, 2005). Acidic, alkaline, oxidative, and photolytic stabilities are required. An ideal stability indicating method is the one that quantifies the standard drug alone and also resolves it from its degradation products (Bakshi, Singh, 2002).

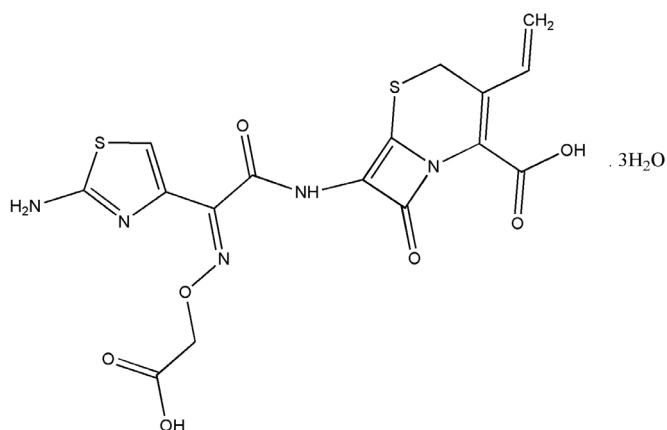


FIGURE 1 - Structure of cefixime trihydrate (CEF).

## EXPERIMENTAL

### Material and reagents

Cefixime trihydrate (CEF) API was available as gift sample from Dr.Reddys Lab, Hyderabad. Methanol, disodium hydrogen phosphate and potassium dihydrogen phosphate were purchased from S.D Fine Chemicals (Mumbai, India) and were of HPLC grade. Hydrochloric acid, sodium hydroxide, hydrogen peroxide and

orthophosphoric acid were purchased from S.D Fine Chemicals (Mumbai, India) and were of A.R grade. HPLC grade water was prepared in house by the triple distillation of water followed by filtration through filter paper of 0.45  $\mu\text{m}$  pore size. The standard solution of 100  $\mu\text{g}/\text{mL}$ , prepared from the API was used for injection onto the HPLC column. The dosage forms used were Cefigip and Taxim-O each of which contained 200 mg per tablet.

### HPLC instrumentation and chromatographic conditions

The HPLC system consists of a Shimadzu LC 20 AD binary pump and SPD 20 A UV detector. The column consists of Phenomenex (250 mm length, 4.6 mm internal diameter, 5  $\mu\text{m}$  particle size) Luna ODS column. LC Solution software was used for data acquisition. Sample injection was done by Rheodyne manual Injector. For the analysis, isocratic mode was selected and the mobile phase is a mixture of methanol and phosphate buffer (pH 6.8) in the ratio of 25:75. Phosphate buffer is prepared with 5.04 g of disodium hydrogen phosphate and 3.01 g potassium dihydrogen phosphate dissolved in 1000 mL of triple distilled water. The mobile phase was pumped at a rate of 1.0 mL/min at room temp. 20  $\mu\text{L}$  of sample was injected each time using rheodyne manual injector. For the detection, the wavelength was set at 298 nm and the peak areas integration was performed using LC solution software.

### Methodology for stress testing

Specificity of the developed method was proved by stress degradation studies where the drug was intentionally degraded under different stress conditions as specified by ICH.

Stress studies were performed by subjecting CEF standard solutions to forced degradation by acidic (1 N HCl), basic (0.1 N NaOH), neutral hydrolytic (purified water) and oxidative degradation (3%  $\text{H}_2\text{O}_2$ ) conditions. A stock solution of 1 mg/mL was prepared in methanol. 2 mL of solution was treated with 2 mL of each of the reagent separately and all the mixtures were kept away from light to exclude the possibility of any photolytic degradation. The acidic mixture was kept for 24 h after which it was neutralised with 1 N NaOH solution and an aliquot was diluted with the mobile phase so as to yield 100  $\mu\text{g}/\text{mL}$ . For the alkaline mixture, prepared with equal volume of 0.1 N NaOH, it was observed that the drug peak disappeared after 30 min, along with many extra peaks indicating that the alkaline degradation was vigorous. Hence the mixture

was kept for 10 min after which it was neutralized with 0.1 N HCl and an aliquot was diluted with the mobile phase to yield 100 µg/mL. Neutral degradation was done in an identical manner using purified water and the mixture was kept for 48 h for the possible degradation, but excluding any neutralization procedure. For oxidative degradation the stock solution was treated with equal volume of 3% H<sub>2</sub>O<sub>2</sub> solution kept for 2 h at room temp after which 100 µg/mL solution was prepared from that for injection.

Photolytic degradation was performed on 100 µg/mL solution, prepared from dry powder exposed to short UV radiations (wavelength = 254 nm) for a period of 24 h.

### Preparation of solutions for injection

10 mg of the reference substance was dissolved in 10 mL of methanol (1000 µg/mL). The working standard solution (100 µg/mL) was prepared by dilution of the stock solution using mobile phase.

A quantity of tablet powder equivalent to 25 mg of CEF was transferred to a 25 mL volumetric flask and added 10 mL of methanol, kept in an ultrasonic bath for 10 min and made upto the volume with methanol and filtered.

### Validation study

The developed method was validated as per ICH guidelines with respect to the following parameters: accuracy, precision, LOD, LOQ, specificity, robustness, stability and system suitability. All the solutions required for validation data were prepared with cefixime standard.

### Linearity

For testing linearity seven calibration standards were prepared in the range of 5 to 100 µg/mL (5, 10, 20, 40, 60, 80 and 100 µg/mL). Standard curve was obtained by plotting peak area against concentration and the evaluation of linearity was done by linear regression analysis using least square method.

### Limit of detection and limit of quantitation

Limit of detection (LOD) and limit of quantitation (LOQ) were estimated at a signal to noise ratio of 3:1 and 10:1 respectively by injecting a series of dilute solutions of known concentration.

### Robustness

Experimental conditions were deliberately altered,

in order to determine the robustness. From the different experimental conditions such as flow rate (1.0 mL/min), lambda max (298 nm) and percentage of methanol (25), each selected factor was changed at three levels (-1, 0, +1). Each factor was changed at a time to study the impact of the change in the experimental conditions on the assay results. Change in the in the peak area and the retention time were noted at for each change in the analytical parameters.

### Stability of sample solution

Sample solution was prepared and analysed by the HPLC instrument using fresh mobile phase at different time intervals (0 h, 8 h and 24 h).

### Accuracy

Accuracy of the developed method was assessed in triplicate at three concentrations (40, 60 and 80 µg/mL). The percentage recovery was calculated from the linear regression equation obtained in the linearity studies.

### Precision

The precision of the analytical method was evaluated by the determination of the repeatability of the method (intra day precision) and intermediate precision (inter day precision) of the sample solutions. Repeatability was calculated by assaying six samples prepared on the same day. Intermediate precision was calculated by assaying 3 days. The relative standard deviation of the area of peaks was calculated.

## RESULTS AND DISCUSSION

### UV spectral analysis

In order to identify the absorption peaks in the UV region, a scan of cefixime trihydrate was taken in methanolic solution of phosphate buffer (25:75), which indicated that 289 nm was the absorption maximum ( $\lambda_{\max}$ ).

### Optimization of chromatographic conditions

To analyze cefixime trihydrate together with its possible degradation products, reverse phase LC in combination with UV detector was developed, optimized and validated. After trials with different mobile phase compositions, a satisfactory separation for the drug and

**TABLE I** - Assay for tablet formulations

| S.NO. | Formulation | Labelled Claim (mg) | Amount found (mg)<br>Mean $\pm$ SD | Assay (%) | %RSD  |
|-------|-------------|---------------------|------------------------------------|-----------|-------|
| 1     | Cefigip     | 200                 | 202.379 $\pm$ 1.983                | 101.19    | 0.984 |
| 2     | Taxim-O     | 200                 | 203.773 $\pm$ 1.657                | 101.89    | 0.813 |

its degradation products were obtained with mobile phase consisting of 10 mM disodium hydrogen phosphate with 0.5% TEA (pH adjusted to 6.3 with OPA) and methanol in the ratio of 75:25 (v/v), pumped at a flow rate of 1 mL/min. The detection was carried out at 289 nm and the retention time was found to be 6.81 min.

### Calibration plot for cefixime trihydrate

A linear calibration plot for the method was obtained over the calibration range of 5 – 100  $\mu$ g/ml at a wavelength of 289 nm. The equation of the regression line was found to be  $y = 84923x + 36508$  with a correlation coefficient of 0.9997. The results showed that an excellent correlation exists between the peak area and concentration of the analyte.

### Analysis of the marketed formulations

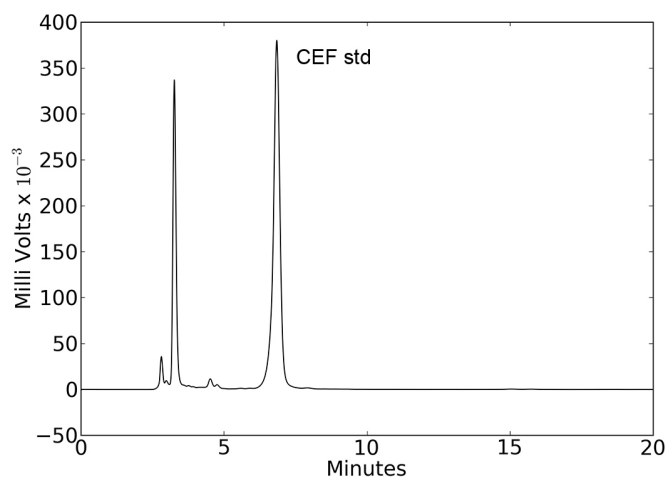
The proposed method was evaluated by the assay of commercially available tablets (formulation-I and formulation- II) containing cefixime trihydrate (200 mg). The results obtained for cefixime trihydrate was compared with the corresponding labeled amounts and reported in Table I. The amount of cefixime trihydrate found in formulation - I (Cefigip) and formulation II (Taxim - O) was 202.37 mg and 203.77 mg respectively. These amounts are within the limits. For both formulation-I and formulation- II the %RSD was less than 2, which indicated the accuracy of the proposed method.

### Stress degradation studies

Cefixime trihydrate was found to degrade under acidic condition (1N HCl). The degradation reaction was more intense and quicker in alkaline condition because the std solution resulted from the alkaline mixture, prepared with an equal volume of 0.1N NaOH, the drug peak disappeared after 30 min, along with many extra peaks. Hence, the reaction time was reduced to 10 min. In the absence of acid or base also, it undergoes hydrolysis, which is revealed by the presence of extra peaks. Upon treatment with 3% v/v H<sub>2</sub>O<sub>2</sub> at room temperature, it was found that

the drug was degraded. Additional peaks were observed after 24 h exposure to UV light indicating that the drug is not photostable.

The chromatograms obtained under different stress conditions are in Figure 2. 1 to 5 and the CEF std drug peak is labelled in all chromatograms. The resolution was found to be more than 2 in all the cases.



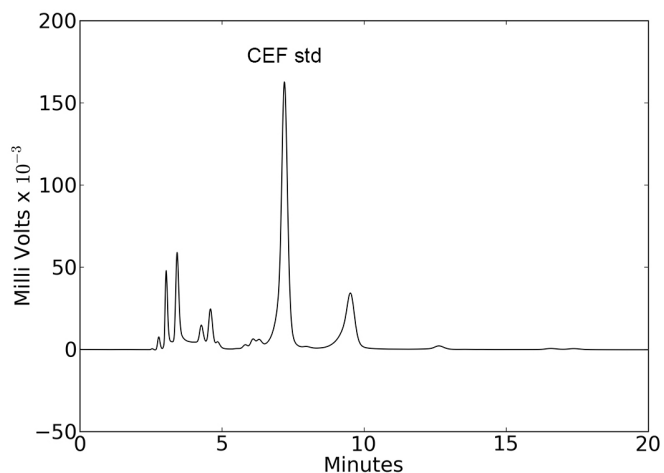
**FIGURE 2-1** - Chromatogram for oxidative degradation (Rt of CEF std = 6.838).

### Method validation

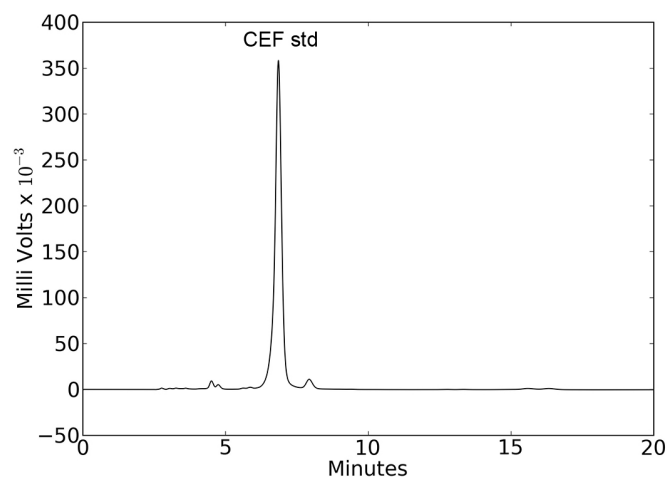
The method was validated for all validation parameters as per ICH guidelines. The linear regression data for the calibration curve ( $n = 3$ ) showed good linear relationship over the concentration range 5-100  $\mu$ g/mL. No significant difference was observed in the slopes of standard curves.

### Precision

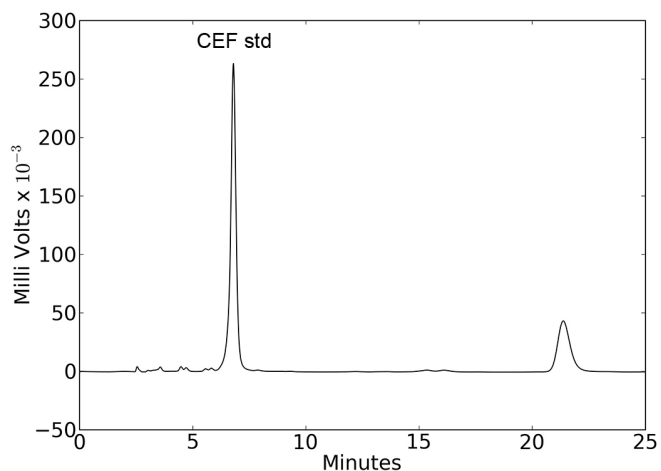
The repeatability (intra-day precision) of the method was determined by intra-day ( $n=3$ ) analysis of three standard solutions of cefixime trihydrate at the concentration of 5, 20 and 40  $\mu$ g/mL. Intermediate precision was determined by the inter-day ( $n=6$ ) analysis of three standard solutions of cefixime trihydrate at the concentration of 5, 20 and 40  $\mu$ g/ml. The data obtained



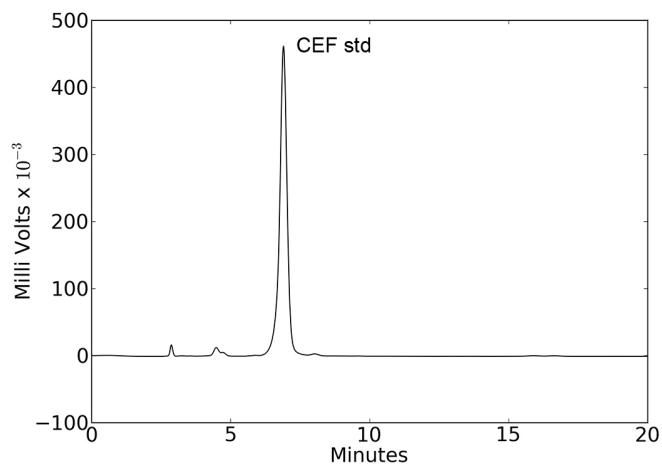
**FIGURE 2-2** - Chromatogram for alkaline degradation (Rt of CEF std = 6.845).



**FIGURE 2-4** - Chromatogram for hydrolytic degradation (Rt of CEF std = 6.91).



**FIGURE 2-3** - Chromatogram for acidic degradation (Rt of CEF std = 6.791).



**FIGURE 2-5** - Chromatogram for photolytic degradation (Rt of CEF std = 6.890).

from precision experiments are given in Table II for intra- and inter-day precision studies. The % RSD values for intra-day and inter-day study were  $< 2.0\%$ , confirming that the method was precise.

### Accuracy (recovery studies)

The proposed method when used for extraction and subsequent estimation of cefixime trihydrate from

**TABLE II** - Data for precision

| Concentration<br>( $\mu\text{g/mL}$ ) | Intra-day Precision (n=3) |       |       | Inter-day Precision (n=6) |       |       |
|---------------------------------------|---------------------------|-------|-------|---------------------------|-------|-------|
|                                       | Amount                    | SD    | %RSD  | Amount                    | SD    | %RSD  |
| 5                                     | 6.029                     | 0.012 | 0.200 | 6.010                     | 0.023 | 0.387 |
| 20                                    | 19.629                    | 0.065 | 0.331 | 19.382                    | 0.303 | 1.567 |
| 40                                    | 39.003                    | 0.465 | 1.192 | 38.837                    | 0.400 | 1.029 |

Acceptance Criteria: % RSD should not be more than 2.

**TABLE III** - Data for recovery studies

| Tablet | Excess drug added to analyte (%) | Theoretical content (mg) | Amount found (mg) | Recovery (%) | SD    | %RSD  | SE    |
|--------|----------------------------------|--------------------------|-------------------|--------------|-------|-------|-------|
| T 1    | 0                                | 20                       | 20.16             | 100.785      | 0.199 | 0.987 | 0.114 |
|        | 80                               | 36                       | 36.51             | 101.416      | 0.132 | 0.362 | 0.076 |
|        | 100                              | 40                       | 40.32             | 100.8        | 0.236 | 0.585 | 0.136 |
|        | 120                              | 44                       | 44.18             | 100.4        | 0.102 | 0.231 | 0.059 |
| T 2    | 0                                | 20                       | 20.31             | 101.54       | 0.165 | 0.810 | 0.095 |
|        | 80                               | 36                       | 36.534            | 101.48       | 0.079 | 0.218 | 0.046 |
|        | 100                              | 40                       | 39.986            | 99.96        | 0.072 | 0.180 | 0.042 |
|        | 120                              | 44                       | 44.386            | 100.88       | 0.354 | 0.797 | 0.204 |

T1, T2: (Cefigip, Taxim-O respectively); Acceptance criteria: % RSD should be <2.

pharmaceutical formulations after spiking with additional drug at three different levels (80%, 100% and 120%) afforded recovery of 99.96–101.54% for two formulations (Cefigip, Taxim-O), and mean recovery and %RSD for cefixime trihydrate from marketed formulations are listed in Table III.

#### Limit of detection (LOD) and limit of quantification (LOQ)

From the linearity plot the LOD and LOQ of cefixime trihydrate were calculated by equation 1 and 2 respectively.

$$\text{LOD} = 3.3 \sigma/S \quad \text{equation 1}$$

$$\text{LOQ} = 10 \sigma/S \quad \text{equation 2}$$

$\sigma$  = standard deviation of the response and S = slope of calibration curve; LOD and LOQ were found to be 0.0398  $\mu\text{g/mL}$  and 0.120  $\mu\text{g/mL}$  respectively which indicate adequate sensitivity of the method.

#### Robustness

In robustness study, each chromatographic factor selected was changed one by one to estimate the effect of change on the results. Thus, replicate injections ( $n = 3$ ) of standard solution at three levels were performed with

small changes in chromatographic parameters (factors). Results, presented in Table IV indicated that the selected parameters remained unaffected by the small variations in these factors. None of the alterations caused a significant change in peak area and tailing factor.

#### System suitability

The system suitability was established by evaluating parameters like theoretical plates, tailing factor, retention time and resolution. The results of system suitability shown in Table V proved that the method is suitable for intended purpose.

#### CONCLUSION

An economical stability-indicating HPLC method has been developed and validated for the determination of Cefixime trihydrate in API and in pharmaceutical formulations. The proposed method is accurate, precise, and specific and also has the ability to separate the drug from degradation products. The degradation products resulting from the forced degradation studies did not interfere with the detection of cefixime trihydrate. The concentration of drug was found to be changing from the initial concentration indicating that cefixime trihydrate undergoes degradation under different stress degradation conditions.

**TABLE IV** - Robustness evaluation

| Parameter  | Retention time | Area (n=3)         | Tailing factor |
|--|----------------|--------------------|----------------|
| <b>A. Flow Rate</b>                              |                |                    |                |
| 0.95   | 7.13           | 8625460            | 0.947          |
| 1  | 6.81           | 8604812            | 0.931          |
| 1.05   | 6.56           | 8662376            | 0.929          |
| Mean ± SD  | 6.83 ± 0.286   | 8630883 ± 29162    | 0.936 ± 0.0098 |
| <b>B. Percentage of Methanol in Mobile Phase</b> |                |                    |                |
| 24   | 7.31           | 8624025            | 0.926          |
| 25   | 6.81           | 8604812            | 0.931          |
| 26   | 6.34           | 8571749            | 0.931          |
| Mean ± SD  | 6.82 ± 0.485   | 8600195 ± 26442    | 0.929 ± 0.0028 |
| <b>C. pH of the Buffer</b>                       |                |                    |                |
| 6.1  | 6.82           | 8669663            | 0.932          |
| 6.3  | 6.81           | 8604812            | 0.931          |
| 6.5  | 6.74           | 8669386            | 0.941          |
| Mean ± SD  | 6.79 ± 0.0436  | 867954 ± 37362     | 0.935 ± 0.0055 |
| <b>D. Wavelength (nm)</b>                        |                |                    |                |
| 287  | 6.89           | 8602180            | 0.942          |
| 289  | 6.81           | 8604812            | 0.931          |
| 291  | 6.83           | 8569139            | 0.931          |
| Mean ± SD  | 6.84 ± 0.042   | 8592044 ± 19879.63 | 0.935 ± 0.0063 |

**TABLE V** - System suitability parameters

| S.No. | Parameters         | Value |
|-------|--------------------|-------|
| 1     | Retention time     | 6.81  |
| 2     | Theoretical plates | 3541  |
| 3     | Tailing factor     | 0.926 |
| 4     | Resolution         | >2    |
| 5     | Capacity factor    | 2.146 |

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