










Co-infection of *Cytauxzoon felis*, *Mycoplasma haemofelis*, and the feline immunodeficiency virus in a domestic cat in Uberlândia, Minas Gerais, Brazil

Co-infecção de *Cytauxzoon felis*, *Mycoplasma haemofelis* e vírus da imunodeficiência felina em gato doméstico em Uberlândia, Minas Gerais, Brasil

Ana Carolina Guimarães Fenelon¹ ; Aline Santana da Hora² ; Karolinn Lourenço da Silva¹ ; Gustavo Henrique Batista de Oliveira¹ ; Marianne Souto e Silva Gonçalves¹ ; Felipe Martins Pastor¹ ; Fernando Cristino Barbosa³ ; Marco Túlio Santos Siqueira¹ ; Fernanda Rosalinski-Moraes⁴ 

¹ Universidade Federal de Uberlândia, Uberlândia – MG, Brazil

² Universidade Federal de Uberlândia, Faculdade de Medicina Veterinária, Laboratório de Investigação Etiológica Veterinária, Uberlândia – MG, Brazil

³ Universidade Federal de Uberlândia, Faculdade de Medicina Veterinária, Uberlândia – MG, Brazil

⁴ Universidade Federal de Uberlândia, Faculdade de Medicina Veterinária, Laboratório de Doenças Parasitárias, Uberlândia – MG, Brazil

ABSTRACT

Cytauxzoon felis is a hemoparasite capable of infecting domestic and wild cats. Studies suggest that wild cats are the main reservoirs of the protozoan, and transmission occurs through the bite of an infected tick. Hemotropic mycoplasmas are Gram-negative bacteria responsible for severe hemolytic anemia. The feline immunodeficiency virus (FIV) is a retrovirus capable of generating immunosuppression in the host and persistent infection. The present work describes a case of co-infection of *Cytauxzoon felis*, *Mycoplasma haemofelis*, and feline immunodeficiency virus (FIV) in a cat. A feline from the rural area was admitted to the Veterinary Hospital of Uberlândia - MG of the Federal University of Uberlândia. Fresh whole blood samples were collected for blood count, enzymatic analysis, DNA extraction, real-time PCR for the detection of *Cytauxzoon felis* and *Mycoplasma haemofelis*, and the Point of Care ELISA test for Feline Leukemia Virus (FeLV) antigens and antibodies to feline immunodeficiency virus (FIV). Piroplasms compatible with *Cytauxzoon* spp. were observed in the smear, and the tests performed were positive for *Cytauxzoon felis*, *Mycoplasma haemofelis*, and FIV.

Keywords: Cytauxzoonosis. Feline retroviruses. FIV. Hemoplasmosis. Mycoplasmosis.

RESUMO

Cytauxzoon felis é um hemoparasita que possui capacidade de infectar felídeos domésticos e selvagens. Estudos sugerem que os felídeos selvagens sejam os principais reservatórios do protozoário e a transmissão ocorre através da picada de um carrapato infectado. Os micoplasmas hemotrópicos são bactérias Gram negativas responsáveis por anemia hemolítica grave. O vírus da imunodeficiência felina (FIV) é um retrovírus com capacidade de gerar imunossupressão no hospedeiro e infecção persistente. O presente trabalho descreve um caso de co-infecção de *Cytauxzoon felis*, *Mycoplasma haemofelis* e vírus da imunodeficiência felina (FIV) em um gato. Um felino proveniente da zona rural foi internado no Hospital Veterinário da Universidade Federal de Uberlândia, em Uberlândia-MG. Foram coletadas amostras de sangue total fresco para a realização de hemograma, análise enzimática, extração de DNA, PCR em tempo real para detecção de *Cytauxzoon felis* e *Mycoplasma haemofelis* e o teste de ELISA *Point of Care* para Antígenos de vírus da leucemia felina (FeLV) e anticorpos para o vírus da imunodeficiência felina (FIV). Em esfregaço foram observados piroplasmas compatíveis com *Cytauxzoon* spp. e os resultados dos testes realizados foram positivos para *Cytauxzoon felis*, *Mycoplasma haemofelis* e FIV.

Palavras-chave: Cytauxzoonose. Retrovíroses felinas. FIV. Hemoplasnose. Micoplasnose.

Correspondence to:

Ana Carolina Guimarães Fenelon
 Universidade Federal de Uberlândia, Campus Glória
 BR-050, Km 78, Bairro Glória
 CEP: 38410-337, Uberlândia – MG, Brazil
 e-mail: ana.cgf@ufu.br

Received: March 31, 2023

Approved: October 27, 2023

How to cite: Fenelon ACG, Hora AS, Silva KL, Oliveira GHB, Gonçalves MSS, Pastor FM, Barbosa FC, Siqueira MTS, Rosalinski-Moraes F. Co-infection of *Cytauxzoon felis*, *Mycoplasma haemofelis*, and the feline immunodeficiency virus in a domestic cat in Uberlândia, Minas Gerais, Brazil. Braz J Vet Res Anim Sci. 2023;60:e210131. <https://doi.org/10.11606/issn.1678-4456.bjvras.2023.210131>.

Cytauxzoon felis is a hemoparasite that can infect domestic and wild felids. Wild felids are believed to be the main reservoirs of the protozoa (Furtado et al., 2017). However, *C. felis* infection in cats has been reported in some regions of the national territory, in the urban areas of large cities (André et al., 2017). The transmission of the agent in the form of merozoites occurs through the bite of an infected tick. The parasite form enters monocytes and macrophages of the host, where they make the first schizogonic cycle. The remaining schizogonic cycles occur in the red blood cells, disrupting the cell (Tarigo et al., 2013). The most common clinical signs are high-grade fever, dehydration, jaundice, anorexia, dyspnea, and pale mucous membranes (Clarke & Rissi, 2015). Diagnosis is through visualization of the merozoites inside the erythrocytes. However, the morphological similarity with other infectious agents confirms the diagnosis via molecular tests (André et al., 2017).

Hemotropic mycoplasmas are round to elongated erythrocytic gram-negative bacteria responsible for severe hemolytic anemia, and *M. haemofelis* is considered the most pathogenic of this group (Sykes, 2010). Although the transmission route is not fully understood, potential vectors are fleas and ticks, and infection through scratching has been reported (Díaz-Regañón et al., 2018; Woods et al., 2005). Cats with hemotropic mycoplasmosis may present with anemia, apathy, pyrexia, inappetence, and mucosal pallor (Tasker, 2010). Polymerase chain reaction (PCR) is currently considered the most sensitive detection method (Messick, 2004; Sykes, 2010).

Feline immunodeficiency virus (FIV) is a retrovirus that promotes persistent infection. The main transmission

route is through bites or injuries from an infected feline. Infection can cause immunodeficiency in cats and, consequently, a propensity for secondary infections, among other complications (Beatty & Sykes, 2021). Rapid tests identifying anti-FIV antibodies diagnose the infection (Little et al., 2020).

Due to the epidemiological importance of recent reports of cytauxzoonosis, this paper describes a case of co-infection of *C. felis*, *M. haemofelis*, and FIV in a domestic cat.

On October 5, 2021, a 10-year-old female castrated feline, 2.600 kg, from the rural area of Uberlândia was referred to the intensive care unit (ICU) of the Veterinary Hospital of the Federal University of Uberlândia. According to the guardian's report, the animal showed no atypical behavior one week before the consultation. Four days prior to the consultation, apathy, ocular hyperemia, and dehydration were noted. On physical examination, the animal was prostrated, 8% dehydrated, anisocoric, and hyperemic with icteric mucous membranes; moreover, an increased volume in the skull region was pertinent. Vital signs showed normoglycemia (176 mg/dL), hypothermia (35.9 °C), and hypotension (86 mmHg).

The complete blood count showed normocytic and normochromic anemia, lymphopenia, and thrombocytopenia (Table 1). A fresh whole blood smear was performed and stained with the Romanowsky panoptic fast method, where piroplasmas were compatible with *Cytauxzoon* spp. (Figure 1) Moreover, eosinophilic structures in cocci, characteristic of hemotropic mycoplasmas inside the red blood cells, were observed. The animal presented hypoalbuminemia, uremia, and hyperbilirubinemia (Table 1). Medications described in Table 2 were prescribed. The animal was referred for overnight hospitalization; however, the guardian opted to care for the patient at home.

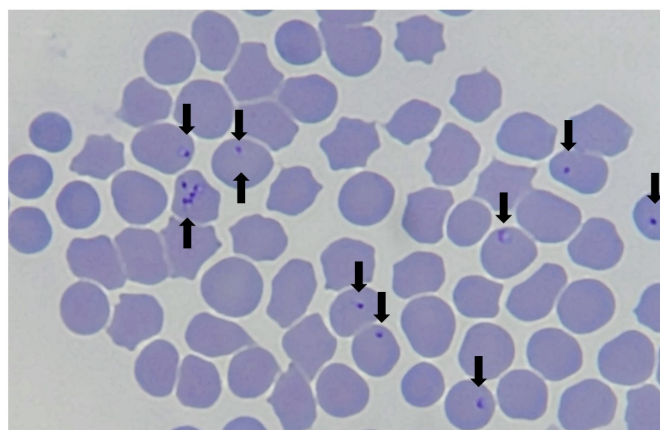


Figure 1 – Blood smear of domestic feline stained by the Romanowsky type method (Fast Panoptic), observing intraerythrocytic structures suggestive of piroplasmids compatible with *Cytauxzoon* spp. (black arrow), 100x magnification in a Nikon Eclipse E200 microscope, Uberlândia – MG.

Table 1 – Hematological and biochemical data of a feline patient from Uberlândia when the diagnosis of infection by *Cytauxzoon felis*, *Mycoplasma haemofelis*, and FIV was obtained

Biochemical and Hematological Parameters	Admission day			Reference values*	
	1 st	2 nd	4 th	Minimum	Maximum
Total red blood cell count (x10 ⁶ µL)	4.08	4.27	-	5.0	10.0
Hemoglobin (g/dL)	6.4	6.7	-	8	15
Hematocrit (%)	17.9	19.8	-	24	45
Mean Corpuscular Volume (fL)	43.9	46.4	-	39	55
Mean Corpuscular Hemoglobin (pg)	15.7	15.7	-	12.5	17.5
Mean Corpuscular Hemoglobin Concentration (g/dL)	35.8	33.8	-	30	36
Plasma Protein (g/dL)	8.4	8.0	-	6.0	8.0
White Blood Cells (mm ³)	10,700	8,100	-	5500	19500
Band Neutrophils (mm ³)	0	405	-	0	300
Segmented Neutrophils (mm ³)	10,272	7,614	-	2500	12500
Eosinophils (mm ³)	0	0	-	0	1500
Basophils (mm ³)	0	0	-	Rare	Rare
Monocytes (mm ³)	214	0	-	0	850
Lymphocytes (mm ³)	214	81	-	1500	7000
Platelets (µL)	36,000	22,000	-	230,000	680,000
Albumin (g/dL)	1.90	-	1.23	2.1	3.3
Alanine aminotransferase (U/L)	72	-	40	6	83
Creatinine (mg/dL)	0.61	-	0.92	0.8	1.8
Alkaline Phosphatase (U/L)	69	-	37	25	93
Urea (mg/dL)	209.6	-	122.6	42.8	64.2
Lactate (mg/dL)	16	-	28	-	-
Total Protein (g/dL)	5.82	-	-	5.4	7.8
Total bilirubin (mg/dL)	21.52	-	24.08	0.15	0.50
Direct bilirubin (mg/dL)	11.31	-	12.93	-	0.20
Indirect bilirubin (mg/dL)	10.21	-	11.15	-	0.20
Calcium (mg/dL)	-	-	5.71	6.2	10.2
Phosphorus (mg/dL)	-	-	6.64	4.5	8.1

*Kaneko et al., 2008; Feldman et al., 2016.

Table 2 – Information on medication administration to the feline patient from Uberlândia - MG when the diagnosis with infection by *Cytauxzoon felis*, *Mycoplasma haemofelis*, and FIV was obtained

Admission Day	Medication administered	Administered dose (mg/Kg)	Route administered	Administration interval
1 st	Omeprazole	10	VO	BID
	Doxycycline	80	VO	SID
	B Complex Vitamins	-	VO	SID
2 nd	Omeprazole	10	VO	BID
	Doxycycline	50	VO	BID
	Dipyron drops	20	VO	SID
	Ondansetron Hydrochloride	4	VO	TID
	Prednisolone	5	VO	BID
	SAME (S-Adenosyl-L-Methionine)	40	VO	SID
	Ursodeoxycholic Acid	30	VO	SID

Caption: VO (oral), SID (once a day), BID (twice a day), TID (three times a day).

On the second hospital day, the animal was returned to the ICU. The guardian reported that the animal did not urinate, defecate, or feed spontaneously during the night. The clinical parameters observed were as follows: icteric mucous membranes, normoglycemia (89 mg/dL), normothermia (38.4 °C), and hypotension (100 mmHg). The animal remained in the ICU and was administered medications described in Table 2.

A new blood count was requested. The results were similar to the first day of hospitalization (Table 1), including the visualization of piroplasmas compatible with *Cytauxzoon* spp. in the ear tip smear and fresh whole blood. The animal was advised for hospitalization for the night. However, the guardian opted for home care, administering the prescribed medications and providing the super-premium adult cat food every 4 h and not

exceeding 25 mL through a nasogastric tube, as prescribed by the manufacturer for a 2.5-kg animal.

On the third hospital day, 300 µL of whole blood sample was collected and submitted for DNA extraction using the commercial Wizard® Genomic DNA Purification Kit (Promega) according to the manufacturer's instructions. After that, the extracted DNA was submitted for real-time PCR for detection of *C. felis* (REF1) and *M. haemofelis*, *Candidatus M. haemominutum*, and *Candidatus M. turicensis* (REF2) using non-specific GoTaq® qPCR Master Mix (Promega) in a Rotor-Gene Q MDx 5 Plex HRM thermal cycler (Qiagen). Feline leukemia virus (FeLV) antigens and FIV antibodies were evaluated by point-of-care enzyme-linked immunosorbent assay test (SNAP® FIV FeLV Combo, IDEXX Laboratories). The test results showed positivity for *C. felis*, *M. haemofelis*, and FIV.

On the fourth hospital day, the animal was again brought with the following clinical parameters: icteric mucous membranes, normoglycemia (123 mg/dL), hypothermia (34.9 °C), and hypotension (120 mmHg). Only biochemical tests and blood transfusion were performed earlier in the day. The biochemical changes included hypoalbuminemia, uremia, and hyperbilirubinemia (Table 1), as per the values available in the reference (Kaneko et al., 2008).

However, due to the significant clinical laboratory alterations and the morbid condition of the animal, euthanasia was offered by the veterinarian, and the guardian authorized the procedure. A necropsy was performed, and the macroscopic alterations found were as follows: icteric skin and mucous membranes, pulmonary and subcutaneous edema, and moderate liver volume increase, with bulging edges and brownish. Multifocal yellowish dots were also observed in the hepatic parenchyma, which deepened on cutting, including a hepatic lobular pattern and renal corticomedullary congestion.

Among the hematological alterations presented by the animal during hospitalization, anemia, lymphopenia, and thrombocytopenia were prominent and congruent with the clinical manifestations Sykes (2010) described in cases of felines infected with hemotropic mycoplasmas. Maia et al. (2013) and Tarigo et al. (2013) reported the presence of lymphopenia, thrombocytopenia, and uremia in *Cytauxzoon* sp.-positive felines, which the animal of the current study manifested. The evolution of hematological alterations that generated an unfavorable prognosis for the animal could be caused by the concomitant infection by different microorganisms.

Among the hemotropic mycoplasmas, *M. haemofelis* is considered the most pathogenic, responsible for severe hemolytic anemia that may culminate in the death of the animal (Messick, 2004). In experimental infections in cats with *M. haemofelis*, Willi et al. (2006) found more severe anemia than in cats infected with other hemoplasma species. Some authors believe that the animal may be an asymptomatic carrier of the agent, and the clinical signs appear when other diseases immunosuppress it (André et al., 2017; Willi et al., 2006). In cats infected with *M. haemofelis*, Firmino et al. (2016) found hematocrit values below the reference value and mild normocytic and normochromic anemia. These data were similar to those presented in this report. The same authors did not find thrombocytopenia in the animals evaluated, contrary to the present study findings.

The hyperbilirubinemia presented by the animal on the first and fourth hospital days was also reported by Maia et al. (2013) study, which was attributed to the infiltration of macrophages with schizonts in the liver and the intense hemolysis caused by *C. felis*. This change can also be found in animals infected with *M. haemofelis*, according to Messick (2004). Contrary to the present findings, Raimundo et al. (2021) found secondary reactive thrombocytosis in an infected feline in Rio de Janeiro.

The presence of icteric mucous membranes and the multifocal yellowish spots in the liver that were reported during necropsy were in line with that reported by Aschenbroich et al. (2012). The same authors further stated that petechial hemorrhages and ecchymoses can be observed throughout the animal's body. However, these changes were not found in the patient. Snider et al. (2010) observed pulmonary edema in cats positive for *Cytauxzoon* sp., also found in the present report. They also noted interstitial pneumonia in cats with cytauxzoonosis, but it was impossible to perform histopathology in this case.

C. felis is an agent known and studied since the 1970s in North America and is associated with fever, apathy, lethargy, jaundice, and high lethality in domestic cats (Rizzi et al., 2015). The same authors report the occurrence of chronic carriers of the parasitosis, with 15 months or more of hemoparasitemia. Brazil has had sporadic reports of *Cytauxzoon* spp. since the early 2000s in domestic (Mendes-de-Almeida et al., 2007) and wild cats (André et al., 2009). André et al. (2022) found approximately 26.15% (n=102/390) positivity for *Cytauxzoon* spp. in the states of Minas Gerais and São Paulo in cats with no changes in clinical pathology examinations. Among these, 63.73% (n=65/102) were from the city of Uberlândia and its neighbor Araguari, which reinforces the agent's presence in this study region.

André et al. (2017) identified two felines positive for cytauxzoonosis in a study conducted in cats in Rio Grande do Norte, without hematological and biochemical changes. These studies support the hypothesis that domestic felines may behave as chronic and asymptomatic carriers of *Cytauxzoon* or that Brazilian isolates are less pathogenic to felines than North American isolates. Additionally, its pathogenicity may be potentiated when associated with other pathogens, generating risks to the animal's health and contributing to an unfavorable prognosis.

Moreira et al. (2018) reported the presence of sheep predation by puma (*Puma concolor*) on the same rural property where the cat of the present report lived, alerting to the possibility of contact of vectors (ticks and fleas) with domestic and wild felines in this location and transmission of infectious agents between species. Due to the absence of work proving how transmission occurs in *C. felis* among felines, some authors believe in the possibility that the jaguar is a reservoir of the protozoan (Furtado et al., 2017). André et al. (2011) found a positive sample, among 123 samples analyzed, for *M. haemofelis* in a wildcat (*Leopardus tigrinus*) without clinical signs of the disease from the region of Jundiá (SP). The cat in the present report may have come in contact with the possible vectors of jaguars or any other wild felid in the area. This could be a source of *C. felis* transmission to the cat. More importantly, since the cat was positive for FIV, wildlife conservation is at risk.

References

André MR, Adania CH, Allegretti SM, Machado RZ. Hemoplasmas in wild canids and felids in Brazil. *J Zoo Wildl Med.* 2011;42(2):342-7. <http://dx.doi.org/10.1638/2010-0198.1>. PMID:22946419.

André MR, Adania CH, Machado RZ, Allegretti SM, Felipe PAN, Silva KF, Nakaghi ACH, Dagnone AS. Molecular detection of *Cytauxzoon* spp. in asymptomatic Brazilian wild captive felids. *J Wildl Dis.* 2009;45(1):234-7. <http://dx.doi.org/10.7589/0090-3558-45.1.234>. PMID:19204356.

André MR, Calchi AC, Furquim MEC, Andrade I, Arantes PVC, Lopes LCM, Demarchi IKLN, Figueiredo MAP, Lima CAP, Machado RZ. Molecular detection of tick-borne agents in cats from Southeastern and Northern Brazil. *Pathogens.* 2022;11(1):106. <http://dx.doi.org/10.3390/pathogens11010106>. PMID:35056054.

The feline patient in this study presented with clinical signs and laboratory alterations compatible with cytauxzoonosis. However, the co-infection with *M. haemofelis* may have contributed to the hematological and biochemical alterations. The presence of FIV may have contributed to the immunocompromised status of the animal, and these two pathogens may have aggravated the infection. Thus, it is impossible to conclude whether *Cytauxzoon* infection would have a benign outcome without co-infection.

This case report demonstrated the presence of *C. felis* in Uberlândia, Minas Gerais, alerting professionals to the possibility of this infectious agent in animals co-infected with other frequent pathogens, such as FIV and *M. haemofelis*. Thus, we encourage the inclusion of cytauxzoonosis in the request for complementary screening tests in the care of cats, preventing the disease from being diagnosed late or completely undiagnosed, as this has an unfavorable prognosis.

Conflict of Interest

There is no conflict of interest.

Ethics Statement

Not applicable.

Acknowledgements

Veterinary Hospital (HOVET) and Etiological Research Laboratory (LIvE Vet) from Federal University of Uberlândia.

André MR, Filgueira KD, Calchi AC, Sousa KCM, Gonçalves LR, Medeiros VB, Ximenes PA, Lelis ICNG, Meireles MVN, Machado RZ. Co-infection with arthropod-borne pathogens in domestic cats. *Rev Bras Parasitol Vet.* 2017;26(4):525-31. <http://dx.doi.org/10.1590/s1984-29612017064>. PMID:29160357.

Aschenbroich SA, Rech RR, Sousa RS, Carmichael KP, Sakamoto K. Pathology in practice: *Cytauxzoon felis* infection. *J Am Vet Med Assoc.* 2012;240(2):159-61. <http://dx.doi.org/10.2460/javma.240.2.159>. PMID:22217023.

Beatty JA, Sykes JE. Feline immunodeficiency virus infection. In: Sykes JE, editor. *Greene's infectious diseases of the dog and cat.* 5th ed. St. Louis: Saunders; 2021. p. 414-28. <http://dx.doi.org/10.1016/B978-0-323-50934-3.00033-1>.

Clarke LL, Rissi DR. Neuropathology of natural *Cytauxzoon felis* infection in domestic cats. *Vet Pathol.* 2015;52(6):1167-71. <http://dx.doi.org/10.1177/0300985814564986>. PMID:25572263.

- Díaz-Regañón D, Villaescusa A, Ayllón T, Rodríguez-Franco F, García-Sancho M, Agulla B, Sainz Á. Epidemiological study of hemotropic *Mycoplasmas* (hemoplasmas) in cats from central Spain. *Parasit Vectors*. 2018;11(1):140. <http://dx.doi.org/10.1186/s13071-018-2740-9>. PMID:29554981.
- Feldman BV, Zinkl JG, Jain NC, Schalm OW. *Schalm's veterinary hematology*. 6th ed. New Jersey: Wiley-Blackwell; 2016.
- Firmino FP, Aquino LC, Marçola TG, Bittencourt MV, McManus CM, Paludo GR. Frequency and hematological alterations of different hemoplasma infections with retrovirus co-infections in domestic cats from Brazil. *Pesq Vet Bras*. 2016;36(8):731-6. <http://dx.doi.org/10.1590/S0100-736X2016000800009>.
- Furtado MM, Taniwaki SA, Metzger B, Paduan KS, O'Dwyer HL, Jácomo ATA, Porfírio GEO, Silveira L, Sollmann R, Tôrres NM, Ferreira Neto JS. Is the free-ranging jaguar (*Panthera onca*) a reservoir for *Cytauxzoon felis* in Brazil? *Ticks Tick Borne Dis*. 2017;8(4):470-6. <http://dx.doi.org/10.1016/j.ttbdis.2017.02.005>. PMID:28196774.
- Kaneko JJ, Harvey JW, Bruss ML. *Clinical biochemistry of domestic animals*. 6th ed. Burlington: Academic Press, 2008.
- Little S, Levy J, Hartmann K, Hofmann-Lehmann R, Hosie M, Olah G, Denis KS. AAFP feline retrovirus testing and management guidelines. *J Feline Med Surg*. 2020;22(1):5-30. <http://dx.doi.org/10.1177/1098612X19895940>. PMID:31916872.
- Maia LMP, Cerqueira AMF, Macieira DB, Souza AM, Moreira NS, Silva AV, Messick JB, Ferreira RF, Almosny NRP. *Cytauxzoon felis* and 'Candidatus *Mycoplasma haemominutum*' coinfection in a Brazilian domestic cat (*Felis catus*). *Rev Bras Parasitol Vet*. 2013;22(2):289-91. <http://dx.doi.org/10.1590/S1984-29612013000200049>. PMID:23856727.
- Mendes-de-Almeida F, Labarthe N, Guerrero J, Faria MCF, Branco AS, Pereira CD, Barreira JD, Pereira MJS. Follow-up of the health conditions of an urban colony of free-roaming cats (*Felis catus* Linnaeus, 1758) in the city of Rio de Janeiro, Brazil. *Vet Parasitol*. 2007;147(1-2):9-15. <http://dx.doi.org/10.1016/j.vetpar.2007.03.035>. PMID:17481822.
- Messick JB. *Hemotropic mycoplasmas* (hemoplasmas): a review and new insights into pathogenic potential. *Vet Clin Pathol*. 2004;33(1):2-13. <http://dx.doi.org/10.1111/j.1939-165X.2004.tb00342.x>. PMID:15048620.
- Moreira TA, Macedo Júnior GL, Souza RR, Bandarra MB, Szabó MPJ. Puma (*Puma concolor*) predating sheep in Uberlândia, Brazil: physical injuries, stress and myopathy. *Biosci J*. 2018;34(3):697-702. <http://dx.doi.org/10.14393/BJ-v34n3a2018-37252>.
- Raimundo JM, Guimarães A, André MR, Baldani CD. *Cytauxzoon felis* DNA detection in healthy cats from Rio de Janeiro, Brazil. *J Parasitol*. 2021;107(5):676-8. <http://dx.doi.org/10.1645/19-159>. PMID:34473290.
- Rizzi TE, Reichard MV, Cohn LA, Birkenheuer AJ, Taylor JD, Meinkoth JH. Prevalence of *Cytauxzoon felis* infection in healthy cats from enzootic areas in Arkansas, Missouri, and Oklahoma. *Parasit Vectors*. 2015;8(1):13. <http://dx.doi.org/10.1186/s13071-014-0618-z>. PMID:25566776.
- Snider TA, Confer AW, Payton ME. Pulmonary histopathology of *Cytauxzoon felis* infections in the cat. *Vet Pathol*. 2010;47(4):698-702. <http://dx.doi.org/10.1177/0300985810364527>. PMID:20442419.
- Sykes JE. Feline hemotropic mycoplasmas. *J Vet Emerg Crit Care*. 2010;20(1):62-9. <http://dx.doi.org/10.1111/j.1476-4431.2009.00491.x>. PMID:20230435.
- Tarigo JL, Scholl EH, Bird DM, Brown CC, Cohn LA, Dean GA, Levy MG, Doolan DL, Trieu A, Nordone SK, Felgner PL, Vigil A, Birkenheuer AJ. A novel candidate vaccine for cytauxzoonosis inferred from comparative apicomplexan genomics. *PLoS One*. 2013;8(8):e71233. <http://dx.doi.org/10.1371/journal.pone.0071233>. PMID:23977000.
- Tasker S. Haemotropic mycoplasmas: what's their real significance in cats? *J Feline Med Surg*. 2010;12(5):369-81. <http://dx.doi.org/10.1016/j.jfms.2010.03.011>. PMID:20417898.
- Willi B, Boretti FS, Baumgartner C, Tasker S, Wenger B, Cattori V, Meli ML, Reusch CE, Lutz H, Hofmann-Lehmann R. Prevalence, risk factor analysis, and follow-up of infections caused by three feline hemoplasma species in cats in Switzerland. *J Clin Microbiol*. 2006;44(3):961-9. <http://dx.doi.org/10.1128/JCM.44.3.961-969.2006>. PMID:16517884.
- Woods JE, Brewer MM, Hawley JR, Wisnewski N, Lappin MR. Evaluation of experimental transmission of 'Candidatus *Mycoplasma haemominutum*' and *Mycoplasma haemofelis* by *Ctenocephalides felis* to cats. *Am J Vet Res*. 2005;66(6):1008-12. <http://dx.doi.org/10.2460/ajvr.2005.66.1008>. PMID:16008224.

Financial Support: None.