

CLINICAL SCIENCE

RADIO FREQUENCY ELECTROMAGNETIC RADIATION (RF-EMR) FROM GSM (0.9/1.8GHZ) MOBILE PHONES INDUCES OXIDATIVE STRESS AND REDUCES SPERM MOTILITY IN RATS

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INTRODUCTION: Mobile phones have become indispensable in the daily lives of men and women around the globe. As cell phone use has become more widespread, concerns have mounted regarding the potentially harmful effects of RF-EMR from these devices.

OBJECTIVE: The present study was designed to evaluate the effects of RF-EMR from mobile phones on free radical metabolism and sperm quality.

MATERIALS AND METHODS: Male albino Wistar rats (10-12 weeks old) were exposed to RF-EMR from an active GSM (0.9/1.8 GHz) mobile phone for 1 hour continuously per day for 28 days. Controls were exposed to a mobile phone without a battery for the same period. The phone was kept in a cage with a wooden bottom in order to address concerns that the effects of exposure to the phone could be due to heat emitted by the phone rather than to RF-EMR alone. Animals were sacrificed 24 hours after the last exposure and tissues of interest were harvested.

RESULTS: One hour of exposure to the phone did not significantly change facial temperature in either group of rats. No significant difference was observed in total sperm count between controls and RF-EMR exposed groups. However, rats exposed to RF-EMR exhibited a significantly reduced percentage of motile sperm. Moreover, RF-EMR exposure resulted in a significant increase in lipid peroxidation and low GSH content in the testis and epididymis.

CONCLUSION: Given the results of the present study, we speculate that RF-EMR from mobile phones negatively affects semen quality and may impair male fertility.

KEYWORDS: Mobile phone; Electromagnetic radiation; Sperm motility; Oxidative Stress; Rats.

INTRODUCTION

Concerns about the possible health effects of mobile phone usage are growing as the number of users has increased tremendously over the past several years. Mobile

phone technology uses radiofrequency electromagnetic radiation (RF-EMR) and has drastically increased the RF-EMR exposure encountered in daily life. Many recent studies have raised questions regarding the safety of such RF-EMR exposure. For example, microwaves generated by mobile phones have been linked to several genetic defects.¹⁻⁵ Research also suggests that microwave radiation from mobile phones may induce chromosomal instability and lead to increased cancer risk.^{3,6,7} Our previous studies have shown that RF-EMR from a 0.9/1.8 GHz mobile phone is anxiogenic (Kumar et al., in press) and impairs avoidance behavior (unpublished data) and spatial memory performance (Sareesh et al., in press) in rats.

As men usually carry mobile phones in their pockets or

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in holders close to their reproductive organs, it is important to evaluate the effects of mobile phone use on male fertility. Although many recent epidemiological studies⁸⁻¹² have suggested that mobile phone use may play a role in male infertility, the mode of action on the male reproductive system remains unclear. Mobile phones might impact the reproductive system via a RF-EMR-specific effect, a thermal effect, or a combination of these effects. In contrast it has recently been suggested that the antioxidant effect of alpha lipoic acid may improve sperm quality.¹³ Therefore, carefully designed studies are needed to properly assess the effects of cell phone use on male fertility.

In the present study, we showed that RF-EMR from a mobile phone in active mode disturbs free radical metabolism in reproductive tissue *in vivo* and thereby leads to changes in semen parameters. Our results suggest a causal relationship between RF-EMR and changes in semen quality.

MATERIALS AND METHODS

Animals: Albino male Wistar rats aged 10-12 weeks were housed in plastic cages (14"×9"×8", 3 rats per cage) inside a well-ventilated room kept at 22±2°C with a 12-hr light:dark cycle. All animals had free access to a standard diet and water. Animals were divided into a control group (group I) and an RF-EMR-exposed group (group II) with 6 rats per group. The Animal Ethics Committee (Manipal University, Manipal) approved all procedures.

Mobile phone exposure: Group II animals were exposed to RF-EMR from a GSM (0.9/1.8GHz) mobile phone in active mode for 1 hour continuously per day for 28 days. To address the concern that any effects of the mobile phone could be due to heat emitted rather than to RF-EMR alone, the phone was kept in a 4"×2"×1" cage with a wooden bottom. Animals were free to move about in the cage during the exposure period and food and water were available *ad libitum*. Control animals were exposed to a mobile phone without battery in a similarly sized cage for the same period in a separate, similar room. Both cages were brought back to the home room following phone exposure.

After the experimental period, all animals were killed by cervical dislocation. The tissues of interest were harvested via incisions in the scrotum and subsequent dissection of the testis and epididymis.

Determination of motile sperm count: The caudal epididymis was excised, placed in a sperm collection vial containing 5 mL BSA-Hanks solution, and cut three times to release the sperm. A 0.05-mL sample of the sperm fluid was removed and diluted with 0.95 mL of BSA-Hanks solution. The number of non-motile sperm in the diluted sperm fluid was determined with a hemocytometer. After the non-motile

sperm had been counted, the vessel containing the diluted sperm fluid was immersed in hot water to kill the motile sperm. The fluid was returned to room temperature and the total number of sperm was counted with the hemocytometer. When gross observation showed a small sperm population in the sperm fluid (low turbidity), an undiluted aliquot of the fluid was used. The percentage of motile sperm was calculated using the following equation:

$$\text{Motile sperm rate (\%)} = \frac{(\text{number of sperm} - \text{number of non-motile sperm})}{\text{Number of sperm}} \times 100$$

Determination of total sperm count: The caudal epididymis used in the above procedure was further cut to release all sperm, and the fluid in the vial was filtered through a nylon-mesh sieve. The resulting stock filtrate was then diluted with formalinized saline (0.1 ml filtrate in 1.9 ml saline), and the number of sperm was determined with a hemocytometer. When the sperm population in the sample was too small (low turbidity), the stock filtrate was not diluted; instead, the vessel was directly immersed in hot water and returned to room temperature before the number of sperm was determined. The measured number of sperm was multiplied by the dilution factor to yield the total sperm count.

Sperm count = Measured number of sperm × Dilution factor

Determination of reduced glutathione (GSH): GSH concentration was determined as described by Cui and Lou¹⁴. Briefly, 10 µL of supernatant from tissue homogenized in 10% TCA (trichloroacetic acid) was mixed with 10 µL of 5,5'-dithio-bis(2-nitrobenzoic acid) (DTNB) in 2.0 mg/2.5 mL methanol. The volume was adjusted to 200 µL with 1.0 M Tris-HCl, pH 8.2 containing 0.02 M EDTA. GSH concentration was quantified by measuring the absorbance of the reaction product at 412 nm (compared to a standard curve of GSH).

Determination of lipid peroxidation (LPO): Oxidative stress induction was indicated by the amount of LPO. LPO was quantified by measuring the formation of thiobarbituric acid reactive substances,¹⁵ expressed as malondialdehyde equivalents (nmol MDA/mg tissue), with 1,1 3,3-tetramethoxypropane used as a standard. After weighing, tissues were homogenized in ice-cold 0.25 M Tris pH 7.4 and centrifuged. The resulting supernatant was mixed with TCA-TBA-HCl [Trichloroacetic acid (TCA): 15% w/v, thiobarbituric acid (TBA) 0.375%, hydrochloric acid (HCl) 0.25 N] and heated for 15 min in a boiling water bath. The lysate was cooled and centrifuged to remove the flocculent precipitate. The absorbance of the supernatant was read at 532 nm.

Student's t tests were performed to determine whether differences between the two groups were statistically significant. The significance threshold was set to $p < 0.05$.

RESULTS

Temperature: Facial temperatures did not deviate by more than 0.1°C after mobile phone exposure in either group. Initial facial temperatures were $32.7 \pm 0.69^{\circ}\text{C}$ in the control group and $32.68 \pm 0.53^{\circ}\text{C}$ in the experimental group. After 1 hr of phone exposure, the average temperatures were $32.9 \pm 0.82^{\circ}\text{C}$ in the control group and $32.83 \pm 0.58^{\circ}\text{C}$ in the experimental group. Temperature differences between the groups were not statistically significant.

Sperm parameters: The total sperm count in controls and RF-EMR-exposed animals were $7.8 \pm 0.21 \times 10^7/\text{mL}$ and $7.6 \pm 0.13 \times 10^7/\text{mL}$, respectively (Fig. 1A). This difference (2.56%) was not statistically significant ($p = 0.052$).

The percent of motile sperm was significantly reduced in RF-EMR exposed animals compared with controls (Fig. 1B). The average percent of motile sperm for controls and RF-EMR exposed animals were $71.97 \pm 8.7\%$ and $43.08 \pm 10.03\%$, respectively ($p < 0.002$), a reduction of $\sim 40\%$.

GSH content: A significant depletion of intracellular GSH was observed in the RF-EMR-exposed group, compared to the control group (Fig. 2A). The decrease was $\sim 10\%$ in the testis and $\sim 24\%$ in the epididymis.

Oxidative damage (LPO): Exposure to RF-EMR from

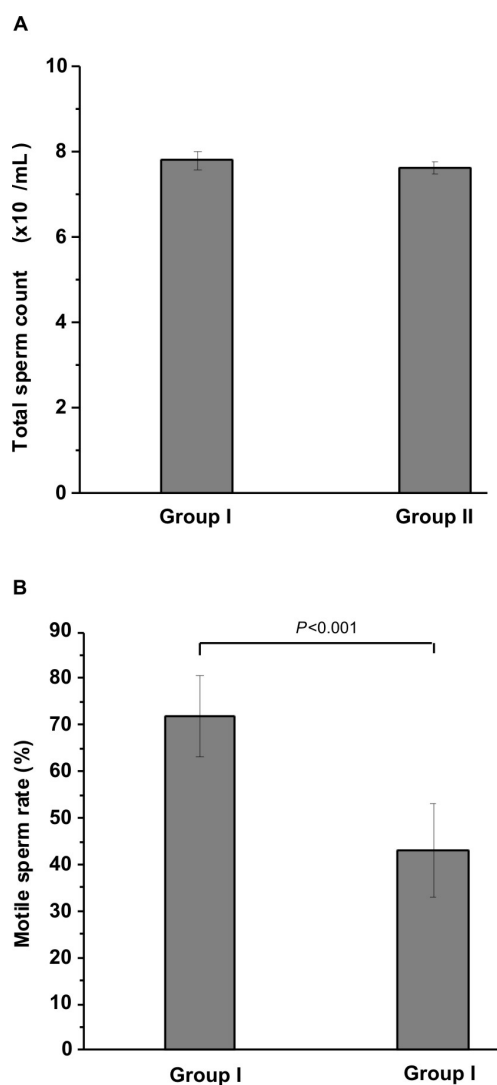


Figure 1 - Sperm count. A. Total sperm count. No significant difference was observed in the total sperm counts for the control (group I) and RF-EMR exposed rats (group II). B. Percent of motile sperm. RF-EMR from a mobile phone significantly reduced the percentage of motile sperm in group II. The results are expressed as the mean \pm standard deviation of six epididymides from six rats.

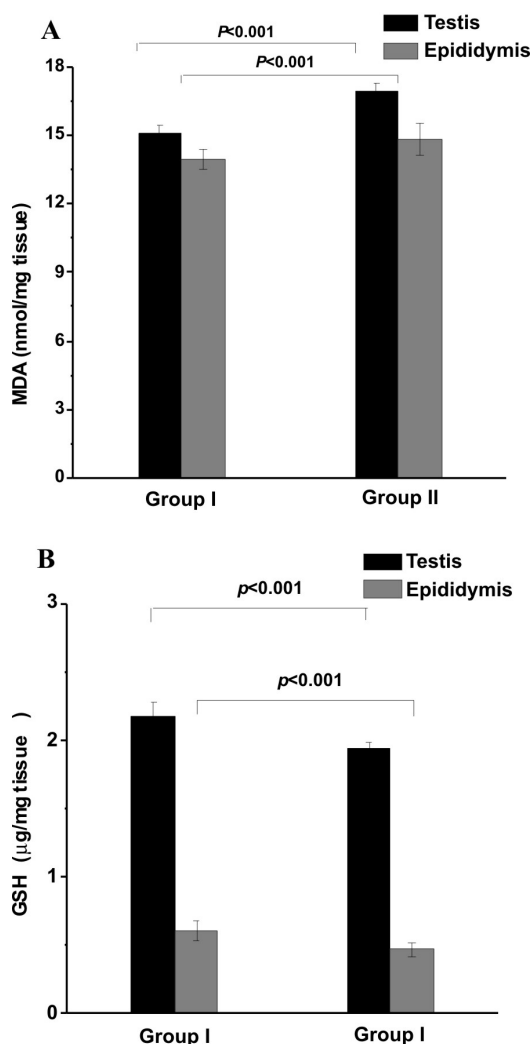


Figure 2 - LPO and GSH. A. Concentration of GSH in reproductive tissues. A significant reduction in testicular and epididymal GSH was observed in rats exposed to RF-EMR (group II) compared to controls (group I). B. Endogenous malondialdehyde (MDA) data. Significant increases were observed in both testicular and epididymal MDA in the rats exposed to RF-EMR (group II) compared to controls (group I). Results are expressed as mean \pm standard deviation of tissue isolated individually from six rats.

the mobile phone increased endogenous MDA levels (an index of *in vivo* LPO) in both the testis and the epididymis (Fig. 2B). The increase was ~8% in testis and ~12% in epididymis.

DISCUSSION

In the present study, we have tested the hypothesis that RF-EMR from mobile phones results in oxidative stress and decreases semen quality. We found a striking reduction in the percentage of motile sperm in rats exposed to RF-EMR, as well as a significantly elevated LPO and decreased GSH content in the testis and epididymis.

Several recent epidemiological studies have investigated the effects of RF-EMR from mobile phones on the human body, but results to date have been contradictory and inconclusive. Furthermore, these studies have not addressed questions regarding the thermal effects of mobile phone exposure.¹⁶⁻¹⁸ Many researchers believe that the effects of mobile phone exposure may be due to the cumulative effects of the heat generated and the RF-EMR emitted from mobile phone; others have suggested that the contribution of the non-thermal component is minimal and that the effects of mobile phone exposure would be negligible if the thermal effect could be eliminated.¹⁹⁻²¹ Certainly, it is widely accepted that temperature increases adversely affect sperm maturity and motility.^{16-18,22,23} In the present study, we excluded temperature effects by keeping the mobile phone in a smaller cage within the home cage. Furthermore, animals were allowed to move freely within the home cage in order to reduce their contact with the phone, and both cages were kept in well-ventilated rooms throughout the experiment to further reduce the possibility of heating. Finally, to confirm the lack of thermal effect, we measured the facial temperatures of rats in both groups both before and after exposure to the phone. After 1 hour of exposure, the mean facial temperature did not differ significantly from the initial temperature in either group. This experimental design eliminated mechanical heat influence from the phone,

and allowed us to conclude that RF-EMR alone affected the reproductive tissue.

Oxidative stress is also a well-established cause of male infertility.²⁴⁻²⁶ Reactive oxygen species (ROS) from spermatozoa and infiltrating leukocytes cause infertility principally by affecting sperm motility.^{27, 28} Spermatozoa possess a multiple plasma membrane redox system that is similar to the transmembrane NADH oxidase. NADH oxidase activity is a major source of superoxide anions²⁹ and, interestingly, RF-EMR has been shown to stimulate NADH oxidase in the plasma membrane of mammalian cells. Normally, ROS is kept at physiologically low levels by intracellular free radical scavengers. GSH, a major thiol in living organisms, is one such scavenger, and plays a central role in coordinating the body's antioxidant defense mechanisms against free radicals. Conditions that perturb intracellular glutathione levels result in significantly altered cellular metabolism. Tissue GSH reflects tissues ability to detoxify, preserve the proper cellular redox balance, and protect cells.³⁰ GSH also probably plays a role in sperm nucleus condensation and spindle microtubule formation.³⁰ The depletion of GSH in RF-EMR exposed animals observed here supports our hypothesis that elevated ROS is responsible for low percentages of motile sperm. Similar findings were reported for human ejaculate exposed to RF-EMR from a mobile phone.³¹ We also believe that RF-EMR induces oxidative stress that affects testicular function and structure in rabbits exposed to mobile phones.³²

Our study validates recent observational studies showing that mobile phone use may play a role in male infertility. Further research will be required to understand the long-term effects of mobile phone use.

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