



## Comments

### Pharmacogenetic testing for thiopurine drugs in Brazilian acute lymphoblastic leukemia patients



In a review article for Clinics in 2018,<sup>1</sup> one of us presented a personal perspective of pharmacogenetic (PGx) testing in oncology in Brazil, highlighting the drug-gene pairs for which there are international consensus guidelines for PGx-informed drug prescription. In 2019, PGx testing for irinotecan:UGT1A1, fluoropyrimidines:DPYD and thiopurines: TPMT/NUDT15 was implemented at Instituto Nacional de Câncer (INCA) in Rio de Janeiro.<sup>2</sup> Here, the authors present results for thiopurines: TPMT/NUDT15, to highlight the development and current status of INCA's PGx program.

PGx testing for thiopurines was designed as an extension of an ongoing project<sup>3</sup> focused on genomic deletions in Acute Lymphoblastic Leukemia (ALL) patients from INCA and nine other oncology centers in Brazil. The study was approved by INCA's Ethics Committee (CAAE 33709814.7.1001.5274), routine procedures for receiving, processing, and storing DNA samples were established and validated allele discrimination TaqMan assays were applied to selected TPMT and NUDT15 single nucleotide polymorphisms (SNPs; [Supplementary Table 1](#)). Selection of the target SNPs, assignment of TPMT, NUDT15, and compound (combined TPMT and NUDT15) metabolic phenotypes according to the TPMT and NUDT15 genotypes, and recommendations for initial thiopurine dosing followed the Clinical Pharmacogenetics Implementation Consortium (CPIC) updated thiopurine guideline.<sup>4</sup>

From the inception of the PGx program until now, 430 ALL patients were successfully genotyped for TPMT and NUDT15 SNPs ([Supplementary Table 2](#)). The frequency distributions of TPMT and NUDT15 alleles,

genotypes and diplotypes, and assigned metabolic phenotypes are shown in [Table 1](#). Minor Allele Frequency (MAF) ranged between 0.012 (NUDT15 rs116855232) and 0.049 (TPMT rs1142345). There were no significant deviations from Hardy-Weinberg equilibrium at any locus, such that homozygosity for the wildtype alleles ranged from 0.907 (TPMT rs1142345) to 0.977 (NUDT15 rs116855232), and heterozygosity varied between 0.023 (TPMT rs116855232) and 0.088 (TPMT rs1142345). Homozygosity for the variant (minor) allele was detected only at the TPMT rs1142345 locus. The allele and genotype frequency of the interrogated SNPs in both TPMT and NUDT15 are in excellent agreement with data from Southeast Brazilian cohorts.<sup>5,6</sup>

The Normal Metabolizer phenotype (NM) was assigned to the vast majority of patients for TPMT (88.4%) and for NUDT15 (97.7%), whereas the Intermediate Phenotype (IM) was assigned to 10.7% (TPMT) and 2.3% (NUDT15) patients. Poor metabolizers (PM) were rare (TPMT, 0.9%) or absent (NUDT15). Four patients (0.9%) were heterozygous carriers of both NUDT15 rs116855232 and TPMT no-function alleles and consequently were assigned the compound IM phenotype, which is associated with lower thiopurine tolerance compared with IMs for either TPMT or NUDT15.<sup>4</sup>

Based on the individual TPMT and NUDT15 combined metabolic phenotypes, four distinct recommendations for thiopurine initial dosing were made ([Fig. 1](#)): i) Start treatment with the usual thiopurine dose: patients with the NM phenotype for both enzymes (86.5%); ii) Consider starting the treatment with reduced thiopurine doses: IMs of either TPMT or NUDT15 (11.6%); iii) Drastic reduction of the initial dose and reduced frequency of administration: PMs: of either or both enzymes (0.9%); iv) Consider a large reduction of the initial thiopurine dosing: compound IMs (i.e. IM for both enzymes) in view of the potential highly increased risk of thiopurine toxicity.

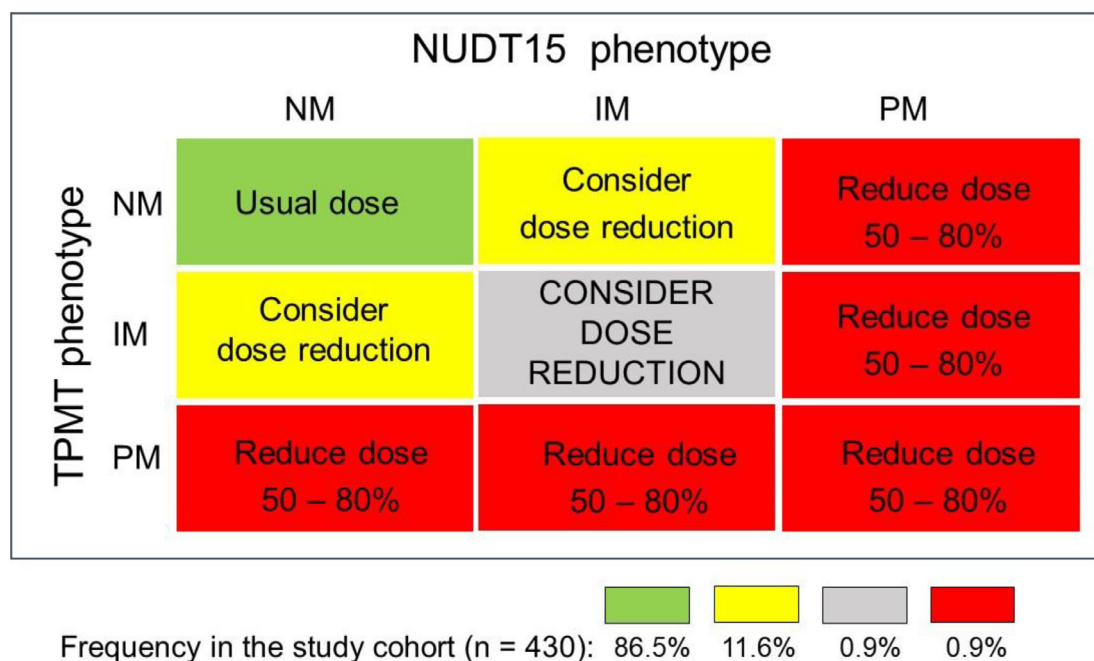
**Table 1**  
TPMT and NUDT15 polymorphisms in Brazilian ALL patients.

Gene / SNPs		Genotype frequency			MAF			
TPMT	wt/wt	wt/var	var/var					
rs1142345	0.907	0.088	0.005	0.049				
rs1800462	0.951	0.049	0.000	0.024				
rs1800460	0.972	0.028	0.000	0.014				
NUDT15								
rs116855232	0.977	0.023	0.000	0.012				
Gene		Diplotype frequency (%)						
TPMT	*1/*1	*1/*2	*1/*3A	*1/*3C	*2/*3*	*2/*3C	*3/*3C	*3C/*3C
	88.4	2.3	4.0	4.4	0.2	0.2	0.2	0.2
Enzyme		Assigned metabolic phenotypes (%)						
	Normal (NM)	Intermediate (IM)	Poor (PM)	Compound Intermediate				
TPMT	88.4	10.7	0.9	-				
NUDT15	97.7	2.3	0.0	-				
Compound	86.5	11.6	0.9	0.9				

wt, Wild-Type allele; var, Variant allele; MAF, Minor Allele Frequency.

<https://doi.org/10.1016/j.clinsp.2023.100214>

Received 13 January 2023; Accepted 25 April 2023



**Fig. 1.** Diagrammatic plot of mercaptopurine dosing recommendation for 430 ALL patients, based on the individual compound metabolic phenotypes for TPMT and NUDT15 enzymes. The recommendations follow the updated CPIC guideline.<sup>4</sup> The proportion of patients receiving each recommendation is shown at the bottom.

The individual TPMT and NUDT15 genotypes and metabolic phenotypes, and the thiopurine dosing recommendation were included in a concise report, which was conveyed by institutional email to the Principal Investigator of each participating institution. The reports cautioned that the dosing recommendation applied to the initial thiopurine dose; subsequent doses should be adjusted based on the degree of myelosuppression and the disease-specific guidelines. The report contained the following disclosure statements: i) The dosing recommendations are based on the CPIC guidelines according to the polymorphisms genotyped; ii) The possibility of influence of other, not interrogated PGx variants cannot be excluded; iii) Adherence to the dosing recommendations is a decision of the prescribing physician.

PGx testing for thiopurines is a landmark in the adoption of PGx as a major instrument for precision (personalized) medicine, and as such led to the creation of the CPIC guidelines.<sup>7</sup> The original thiopurine guideline contemplated only TPMT polymorphisms; the inclusion of NUDT15 polymorphisms in the 2018 updated guideline attests to the dynamics of PGx research but also highlights the challenge that dosing recommendations must be updated as novel evidence of PGx associations emerges. Implementation of PGx testing at INCA to inform the prescription of mercaptopurine to ALL patients is a pioneering initiative within the Brazilian Public Health System, which hopefully will prompt similar programs in other medical centers in Brazil, whether public or private. Financial support from the Brazilian agencies Decit/MS, FAPERJ, and CNPq, and the commitment of a dedicated task force at INCA were decisive factors for the success of the program and assured its continuity throughout the COVID-19 pandemic.

#### Conflicts of interest

The authors declare no conflicts of interest.

#### Acknowledgments

The authors are grateful to André Francisco Duarte and Alessandra Faro de Jesus for their excellent technical support. The authors are also grateful to Dr. Marcelo Land and Dr. Elaine Sobral da Costa for their continuous collaboration with the ALL Project. The PGx program at INCA receives financial support from the Brazilian agencies CNPq (Conselho

Nacional de Desenvolvimento Científico e Tecnológico) DECIT/MS (Departamento de Ciência e Tecnologia/Ministério da Saúde) and FAPERJ (Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro).

#### Supplementary materials

Supplementary material associated with this article can be found in the online version at [doi:10.1016/j.clinsp.2023.100214](https://doi.org/10.1016/j.clinsp.2023.100214).

#### References

- Suarez-Kurtz G. Pharmacogenetic testing in oncology: a Brazilian perspective. *Clinics* 2018;73(suppl 1):e565s.
- Suarez-Kurtz G, Kovaleski G, Elias AB, Motta V, Wolch K, Emerenciano M, et al. Implementation of a pharmacogenomic program in a Brazilian public institution. *Pharmacogenomics* 2020;21(8):549–57.
- Maciel ALT, Barbosa TDC, Blunck CB, Wolch K, Machado AAL, da Costa ES, et al. IKZF1 deletions associate with CRLF2 overexpression leading to a poor prognosis in B- cell precursor acute lymphoblastic leukaemia. *Transl Oncol* 2022;15(1):101291.
- Relling MV, Schwab M, Whirl-Carrillo M, Suarez-Kurtz G, Pui CH, Stein CM, et al. Clinical pharmacogenetics implementation consortium guideline for thiopurine dosing based on TPMT and NUDT15 genotypes 2018 Update *Clin Pharmacol Ther* 2019;105(5):1095–105.
- Reis M, Santoro A, Suarez-Kurtz G. Thiopurine methyltransferase phenotypes and genotypes in Brazilians. *Pharmacogenetics* 2003;13(6):371–3.
- ABraOM: Brazilian genomic variants [Accessed December 15, 2022]. Available from <https://abraom.ib.usp.br/>.
- Relling MV, Klein TE, Gammal RS, Whirl-Carrillo M, Hoffman JM, Caudle KE. The clinical pharmacogenetics implementation consortium: 10 years later. *Clin Pharmacol Ther* 2020;107(1):171–5.

Guilherme Suarez-Kurtz<sup>a,\*</sup>, Cristina Wiggers Almeida<sup>b,c</sup>, Eduardo Chapchap<sup>d</sup>, Márcia Trindade Schramm<sup>e,f</sup>, Maura Rosane Valério Ikoma-Coltutato<sup>g,h</sup>, Mecneide Mendes Lins<sup>i</sup>, Teresa Cristina Cardoso Fonseca<sup>j</sup>, Thais Ferraz Aguiar<sup>k</sup>, Mariana Emerenciano<sup>l,\*</sup>

<sup>a</sup> Divisão de Pesquisa Clínica e Desenvolvimento Tecnológico, Instituto Nacional de Câncer, Rio de Janeiro, RJ, Brazil

<sup>b</sup> Hospital Federal da Lagoa (HFL), Rio de Janeiro, RJ, Brazil

<sup>c</sup> Hospital Universitário Pedro Ernesto, Universidade do Estado do Rio de Janeiro (UERJ), Rio de Janeiro, RJ, Brazil

<sup>d</sup> Hospital Albert Einstein, São Paulo, SP, Brazil

<sup>e</sup> Hospital do Câncer I, Instituto Nacional de Câncer, Rio de Janeiro, RJ, Brazil

<sup>f</sup> Prontobaby Hospital da Criança Ltda, Rio de Janeiro, RJ, Brazil

<sup>g</sup> Fundação Amaral Carvalho, Jaú, SP, Brazil

<sup>h</sup> Sabin Medicina Diagnóstica, Brasília, DF, Brazil

<sup>i</sup> Instituto de Medicina Integral Professor Fernando Figueira, Recife, PE, Brazil

<sup>j</sup> Hospital Manoel Novaes-Santa Casa de Misericórdia de Itabuna, Itabuna, BA, Brazil

<sup>k</sup> Instituto Estadual de Hematologia Arthur Siqueira Cavalcanti, Rio de Janeiro, RJ, Brazil

<sup>l</sup> Divisão de Pesquisa Básica e Experimental, Instituto Nacional de Câncer Rio de Janeiro, RJ, Brazil

\*Corresponding author.

E-mail address: [kurtz@inca.gov.br](mailto:kurtz@inca.gov.br) (G. Suarez-Kurtz).