

Gene therapy: what it is, what it is not and what it will be

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Introduction

FROM ITS foundation by the monk Johann (Gregor) Mendel in the nineteenth century to date, genetics has evolved dramatically and earned an important place among the sciences. The sequencing of the human genome (Lander et al., 2001, Venter et al., 2001), a great achievement that promises to accelerate the progress of biology and medicine in the twenty-first century, was completed ten years ago.

Every day modern medicine makes important discoveries in research areas aimed at developing new paradigms for treating diseases that are still incurable. Among them, the expectation of curing genetic diseases rests on the identification of genes responsible for its pathogenesis and on the advancement of recombinant DNA technology or “genetic engineering”, which enable manipulating the genome in an increasingly efficient and safer way (Watson et al. 2006). In parallel, the determination of genetic susceptibility to certain diseases, the course and clinical manifestations thereof (NCBI, 2009), as well as the tremendous advance in understanding the cellular and molecular biology of fundamental pathological events such as inflammatory processes, proliferative disorders and programmed cell death (Tsongalis & Coleman, 2009), all increase the expectation that genome manipulation may someday be applied to a wide range of diseases.

This is still an incipient area of medicine, practiced especially in fundamental research laboratories, and its application is still strictly experimental. Although there area in this area commercial products approved for medical use (Pearson et al., 2004), the expectation of scientists, as well as of the pharmaceutical and biotechnology industry, is that the release of genome manipulation protocols for medical practice and the respective market of biological products should advance cautiously over the next 5-10 years, but still limited to a restricted number of applications.

In 1990, however, a medical team in the U.S. inserted a healthy gene in the body of a sick girl, whose health improved after the treatment. A new era had begun. The era of gene therapy, that is, the procedure of introducing healthy genes into an organism using recombinant DNA techniques (in this

context called “therapeutic genes”) to replace, manipulate or supplement inactive or dysfunctional genes (Linden, 2008).

The beginning of gene therapy

From the 1940s genetics gained great momentum, and discoveries about the nature, chemical composition and properties of the genetic material, as well as the first manipulations of the DNA of bacteria began to generate expectations of novel therapeutic advances.

The mid-1960s were marked by speculation about the possibility of using viruses to transfer genes to human patients and cure genetic diseases (Friedmann, 1997). Back then, scientists already believed that the genes of certain viruses could be effective and that it was possible to insert healthy human genes in viruses which, in turn, would transfer them to the patient. However, it was only in the beginning of the following decade that Paul Berg succeeded in actually manipulating a DNA molecule (Jackson et al., 1972), creating the recombinant DNA technology.

Two initial attempts to apply the gene therapy concept to clinical practice failed, one for relying on an assumption about the properties of a virus, which later proved to be false (Rogers, 1952; Rous & Rogers, 1951; Andrewes, 1966; Friedman, 2001; Scaglia & Lee, 2006); the other, although technically justifiable and already using recombinant DNA methodologies, was marred by a serious ethical lapse (Mercola & Cline, 1980). But in 1989 a new test conducted in accordance with the rules in force at the time restored positive expectations in this research area.

The patient treated in 1989 was a four-year-old girl who had been deprived of a normal life because of a genetic disorder caused by a deficiency of the adenosine deaminase enzyme (ADA), which is essential for the development of the immune system. Several mutations in the gene encoding the enzyme cause ADA deficiency, which results in degeneration of the T cells of the immune system (Buckley, 2004) and is one of the main causes of severe combined immunodeficiency syndrome (SCID). In this case, the disease is known by the acronym ADA-SCID. Children affected by various forms of SCID (*ibid.*) have very low resistance to infection and, if untreated, usually die before six months of age. They are known as “bubble children” because they need to be isolated, often in a plastic bubble. Treatment usually entails replacing the enzyme with weekly injections. In the case in question, after a period of one year of relative success, in the second year of treatment the child was again plagued by frequent infections and developed an allergy to the enzyme preparation used for injections, indicating that the enzyme replacement therapy was failing. Dr. William French Anderson, from the University of Southern California, was then authorized by the ethics committees to initiate a clinical gene therapy trial (Anderson et al., 1990).

Every one or two months researchers took T cells from Ashanti’s blood,

inserted the ADA gene, induced the proliferation of these cells in the laboratory and then re-infused the treated cells into the patient's blood stream (Culver et al. 1,991). After seven infusions there was a six-month break, and then the infusions were resumed up to two years of treatment. For safety purposes, the girl continued to receive weekly injections of the enzyme. Gene therapy in this patient, as well as that started in 1991 in a second nine-year old patient, yielded positive results. There was clinical improvement with a reduction in the amount of enzyme that needed to be replaced. It was observed that enzyme levels in the patients' blood increased progressively as a result of the gene therapy and remained stable during the six-month break period (Blaese et al. 1995; Mullen et al. 1996). Finally, twelve years after the end of the infusions, when the two cases were reevaluated, large numbers of T cells continued to express the therapeutic gene in the first patient's blood, whose treatment was more successful than that of the second girl (Muul et al. 2003).

It should be noted that there are still technical issues related to this study that prevent us from considering it a complete clinical success. As the children continued to receive enzyme replacement, although in smaller doses, there is cause for doubt about how much the gene therapy has actually contributed, for example, for the first patient, now 24 years old, to be leading a healthy and active life. However, since the treatment of these first two patients, gene therapy for ADA-SCID has evolved and today is considered a clinical success (Aiuti et al., 2009; Candotti & Kohn, 2009). Although in its infancy, the study started in 1989 and which has produced at least some positive results in compliance with ethical requirements, is a milestone in the history of gene therapy and has inspired the subsequent growth of this area of scientific research.

Forms of gene therapy

The idea of using recombinant DNA techniques to repair the genome was inspired by diseases caused by mutation in a single gene (the so-called monogenic diseases). In this case, the idea is to replace or supplement dysfunctional gene expression by inserting one or more copies of the therapeutic gene (Porteus et al. 2,006; O'Connor & Crystal, 2006; Brinkman et al. 2006). The treatment of ADA-SCID is a successful application of this idea.

But monogenic diseases are not the only target of gene therapy (Figure 1). Modern medicine fights many complex diseases, whose primary causes are not yet known and for which there is, at best, only palliative treatments. In certain cases, it is possible to plan a gene therapy intervention to reduce or prevent progression of the disease. The intervention can be based on the knowledge of genetic determinants of susceptibility or severity, or on the opportunity for changing fundamental mechanisms or the physiology of cells, organs or systems affected by the diseases (Cardone, 2007; Flotte, 2007). The main strategies involve increasing cellular resistance, stimulating regeneration or repair systems, or restoring functional characteristics specific to certain organ systems

by modulating genes not necessarily related to the cause of the disease (Bagley et al., 2008; Lundberg et al. 2008). In the case of tumors, the main objective is the selective induction of cell death in proliferating cell populations (Bauzon & Hermiston, 2008; Cattaneo et al., 2008; Ribacka et al., 2008).

Finally, there is a peculiar form of gene therapy called DNA vaccine, in which instead of using a protein or a whole inactivated virus, as is the case in conventional vaccines, the patient receives the gene encoding a protein typical of the aggressor agent. The patient's body will then begin to permanently produce the exogenous protein, stimulating its own immune system. These vaccines can have a preventive purpose, similarly to classical vaccines, or a curative purpose, leading the immune system to attack the aggressor agents already established in the body (Atkins et al., 2008 Sykes, 2008; Silva et al., 2009).

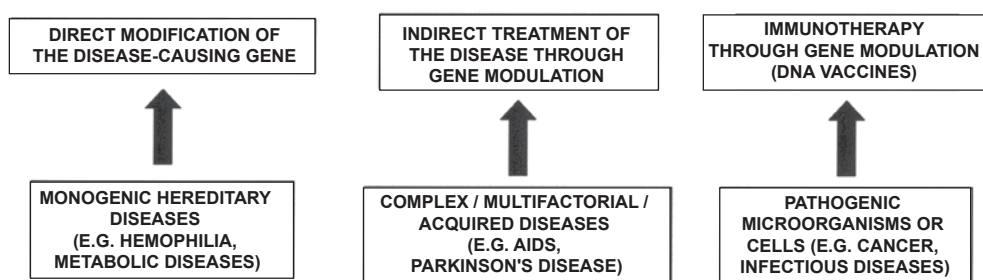


Figure 1 – Main forms of gene therapy

Cell therapy, stem cells and gene therapy

Stem cells are currently the most-commonly referenced medical topic in the media. At the same time, there is some confusion as to the actual meaning of the terms stem cells, cell therapy and gene therapy. In the so-called cell therapy, whole cells are used to treat an illness, based on the regenerative properties of stem cells or on other effects - most of which have not yet been explained - of the transplanted cells. The classic example, whose foundation is well known, is leukemia, but there is expectation that many classes of diseases will be treated with the use of cell therapy in the coming years (Torrente & Polli, 2008; Gribben, 2008; Einstein & Ben-Hur, 2008; Reffellmann et al., 2008).

In this context, it is important to note that cell therapy does not necessarily involve genetic modification. Gene therapy, in turn, is based on the introduction or modification of genes. This can be done directly in vivo, without the aid of whole cells from the patient or from a donor.

That is, gene therapy and cell therapy are two different concepts. However, there are methods that combine the two techniques. An example of combined gene therapy and cell therapy was again the aforementioned ex vivo procedure that started gene therapy. Novel gene therapy technologies for ADA-SCID are

based on the genetic manipulation of bone marrow-derived stem cells, instead of the T cells used in the initial studies (Aiuti et al. 2009). Therefore, under certain circumstances cells can be used as a vehicle to introduce the therapeutic gene. However, it is the introduction of genes and the use of recombinant DNA technologies that characterize a treatment as gene therapy.

Vectors for gene therapy

The basis of gene therapy lies in delivering genes to cells. However, the entry of pure DNA through the plasma membrane of eukaryotic cells is extremely rare (Vellai & Vida, 1999). This difficulty, of course, is beneficial for the body, as it hinders spurious changes in cellular metabolism and even changes similar to those observed in the evolution of species.

Therefore, in general, a carrier is needed to facilitate the delivery of DNA to living cells. This carrier is called “vector”. There are three main classes of vectors currently under development: plasmids, viral vectors and nanostructured vectors.

Plasmids

Plasmids are relatively simple DNA sequences, but effective for gene expression, in which it is possible to deliver a therapeutic gene using recombinant DNA techniques (Voss, 2007; Clanchy & Williams, 2008; Gill et al., 2009). But breaking the resistance of cells to the introduction of plasmids requires weakening the cell membrane, which can be done by various methods such as the use of electroshock or chemical substances that chemically weaken the cell membrane (Dass, 2004; & Cemazar & Sersa, 2007; Favard et al., 2007; Wu & Lu, 2007). Another alternative is to apply a large number of plasmids in the vicinity of the cells so that, even with very low efficiency, a small fraction that succeeds in crossing the membrane will be effective, or, still, to rapidly inject a large amount of a solution containing plasmid (Herweijer & Wolff, 2007).

These techniques are, however, very limited. For example, it is unlikely that they will ever be used to deliver genes to organs of difficult access like the brain. Thus, the use of plasmid vectors is limited to certain cases, such as delivery by intramuscular injection, in the case of DNA vaccines, or in the cardiac muscle, or still in experimental studies in animals. Nonetheless, this technology can have important applications such as, for example, to deliver the healthy gene to isolated cells and to combine gene therapy with cell therapy.

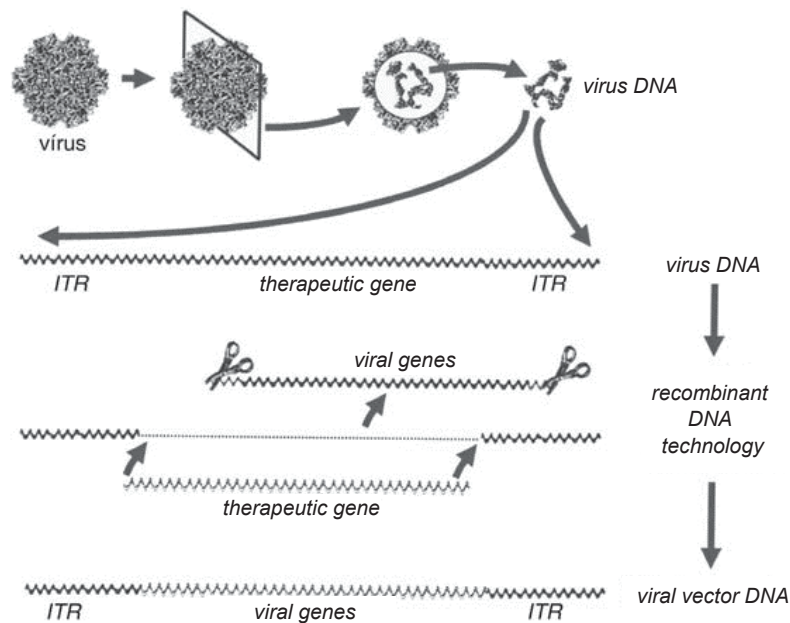


Figure 2 – Construction of a viral vector for gene therapy. The figure illustrates, as an example, the adeno-associated virus, whose genetic material is a single-stranded DNA. At the top is the scheme showing the outside and inside of a virus, in a section showing the location of viral DNA. This DNA contains several genes necessary for the life cycle of the virus, i.e., its multiplication and rearrangement inside the cells. But to be used as a vector, the DNA of terminal regions, identified by the acronym ITR, is enough. The process consists of replacing the viral genes with the therapeutic gene, using recombinant DNA technology. The essential component of the viral vector is therefore produced. However, as naked DNA does not enter the cells easily, it is necessary to reassemble a virus similar to the one shown in the upper left corner of this figure and in large quantities, as illustrated in Figure 3. Reproduced from Linden (2008), with permission of the publishers.

Viral vectors

In contrast to the resistance of the cell membrane to the spontaneous entry of DNA into a cell, viruses are microorganisms specialized precisely in invading cells and introducing genetic material therein. They contain nucleic acid (DNA or RNA) surrounded by a layer of protein and, in some cases, by an additional envelope of proteins and lipids, and their life cycle involves releasing viral nucleic acid inside the host cell. This property is exploited for delivering therapeutic genes to cells using recombinant DNA technologies.

Some vectors are derived from adenovirus. This family includes almost 50 different types of viruses that cause, for example, pharyngitis or conjunctivitis. Infections by adenovirus are very common and, therefore, the majority of the population has antibodies against one or more types of this virus family. Others

are members of the retroviruses family, which includes the HTLV that causes a type of leukemia and the HIV that causes AIDS, a member of the lentiviruses subfamily, which have been widely studied as a source of vectors for gene therapy. There are also other vectors derived from viruses of the adenovirus-associated family, which are not pathogenic to humans.

The principle of production of viral vectors for gene therapy (Figs. 2 and 3) consists in removing the genes involved in pathogenic and viral proliferation mechanisms and keeping only the necessary for the invasion of cells without multiplication, followed by the introduction of a therapeutic gene into the remaining viral DNA (Machida, 2002). Removing genes responsible for the pathogenic character and for multiplication enables, for example, a virus of the same subfamily of the dangerous HIV to give rise to a viral vector useful for gene therapy.

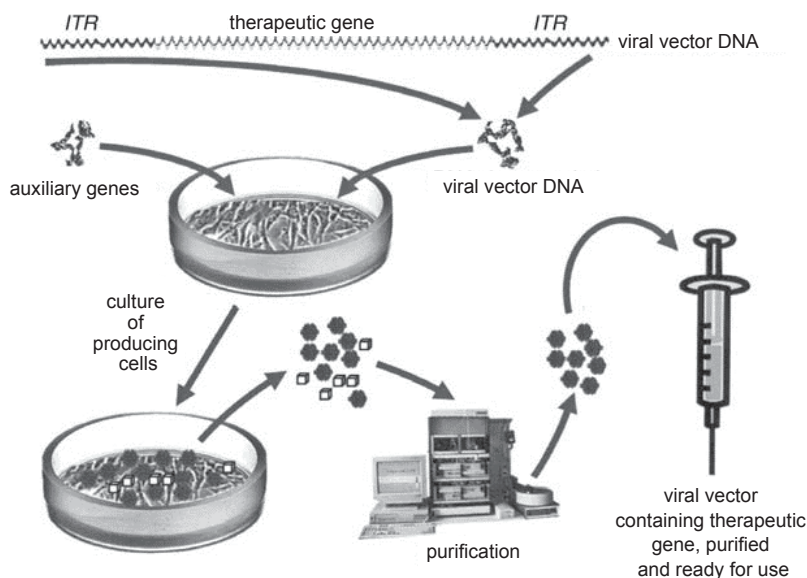


Figure 3 – Mass production of viral vectors for gene therapy. The figure once again uses as an example a vector derived from adeno-associated virus. The viral vector DNA was constructed as shown in Figure 2. This DNA is introduced by precipitation or electroporation into producing cells together with a plasmid containing auxiliary genes, which are required for packaging the vector DNA into the structure of viruses similar to the original adeno-associated viruses. Producing cells form large quantities of complete viral vectors together with contaminants, which are removed at a purification stage, after which trillions of viral particles containing the therapeutic gene free of impurities are obtained. The vector is therefore ready for use. Reproduced from Linden (2008) with permission of the publishers.

Viral vectors differ from one another (Table 1). Some are more efficient while others are more capable of carrying large genes. Some are more likely to

cause inflammatory reactions than others. Finally, some vectors, such as those derived from retroviruses, have the property of integrating into the cell genome. This is positive when one wants a permanent expression of the therapeutic gene; however, it may cause serious adverse effects.

Table 1 – Properties of various types of viral and non-viral vectors for gene therapy

	Retrovirus	Lentivirus	Herpesvirus	Adenovirus	Adeno-associated	Plasmid	Nano-structured
Provirus	RNA	RNA	RNA	DNA	DNA	DNA	DNA or RNA
Capacity	~ 9 kB	~ 10kB	> 30 kB	~ 30 kB	4.6 kB	unlimited	variable
Integration into the recipient's genome	yes	yes	yes	no	extremely rare	no	no
Transgene rearrangements	+	-	-	-	-	-	-
Duration of transgene expression	long	long	transient	transient	long in postmitotic cells	transient	transient
Transduction of postmitotic cells	-	+	+++	+++	++	+	+
Immunity pre-existing in recipient	no	no	yes	yes	yes	no	no
Adverse effects	insertional mutagenesis	insertional mutagenesis	inflammatory response	inflammatory response	mild inflammatory response	no	?
Germline transmission	-/+	+	-	-	-/+	-	?

Source: Modified from Nathwani et al. (2005).

Nanostructured vectors

Another way to introduce DNA into cells is being developed from preparations obtained using advanced nanotechnology techniques (Sanvicens & Marco, 2008). This includes polymers that form real networks that hold a gene and release their load when they penetrate the cells, as well as lipid vesicles containing DNA, which are capable of fusing with the cell membrane, releasing their contents inside the latter.

These vectors can be enriched with molecules that help to specify into which types of cells the content can penetrate, or that enable guiding or selectively transferring the vectors from one compartment to another, e.g., from the blood to the brain (Pardridge, 2005, 2007 Figure 4). This technique is important because it will enable brain gene therapy without the need for neurosurgery to introduce the vector, since intravenous injections would suffice.

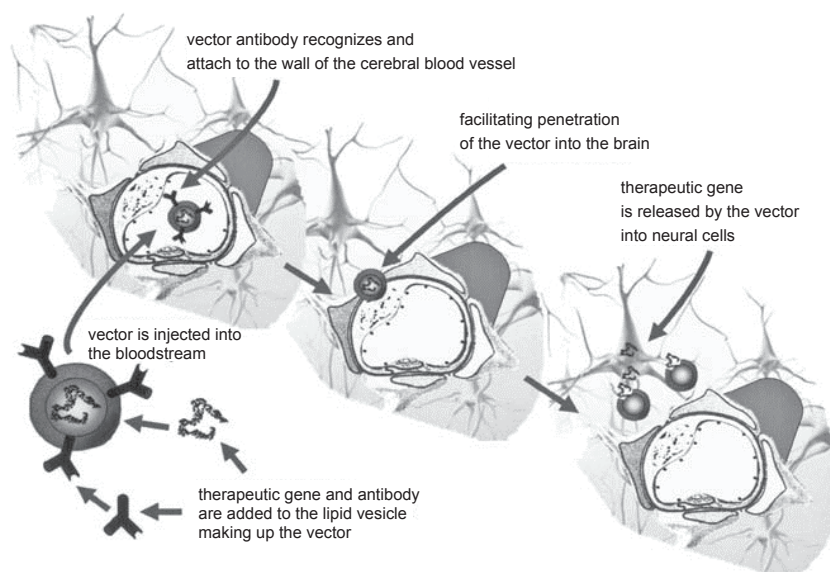


Figure 4 – Model of non-viral vector combined with a routing molecule. The figure uses as an example an immunoliposome for cerebral gene transduction (Pardridge, 2005). As the wall of cerebral blood vessels is highly resistant to drug penetration, including DNA, from the blood, the vector is composed of a vesicle formed by lipids containing DNA with the therapeutic gene. Antibodies against the transferrin receptor, which recognize this receptor on the surface of cells of the wall of cerebral blood vessels and of neurons are inserted on the surface of the vesicle. So, when the vector is injected into the blood stream, it attached strongly to the wall of cerebral blood vessels, thus facilitating its penetration into the brain tissue and, consequently, the introduction of the therapeutic gene into neural cells. This technique can be used to route vectors to the appropriate destination, based on the choice of the antibody inserted on the surface of the vector, which should be selective for blood vessels of the organ to be treated. Reproduced from Linden (2008) with permission of the publishers.

In other cases, cells modified by the insertion of a therapeutic gene can be encapsulated in compartments produced from inert polymers and then introduced into the body. The advantage of this technique is that the cells can produce and secrete therapeutic molecules while isolated from the patient's immune system (Hauser et al. 2004; Lindvall & Wahlberg, 2008). Therefore, encapsulated cells do not need to be derived from the patient him or herself.

Gene therapy today

Gene therapies are novel procedures still in the experimental stage. Basic knowledge has been acquired in fundamental research laboratories through tests in experimental models and preclinical trials. These studies validate the potential efficacy of a therapeutic strategy, and enable detecting potential risks to humans, anticipating changes in vectors and other components of the therapeutic strategy that increase safety for human use.

Fundamental research in gene therapy is intense and growing in the world. Figure 5 illustrates the continued growth in the number of scientific publica-

tions in this area. In the last three years, about 30 scientific papers, on average, have been published *each day* on topics related to gene therapy.

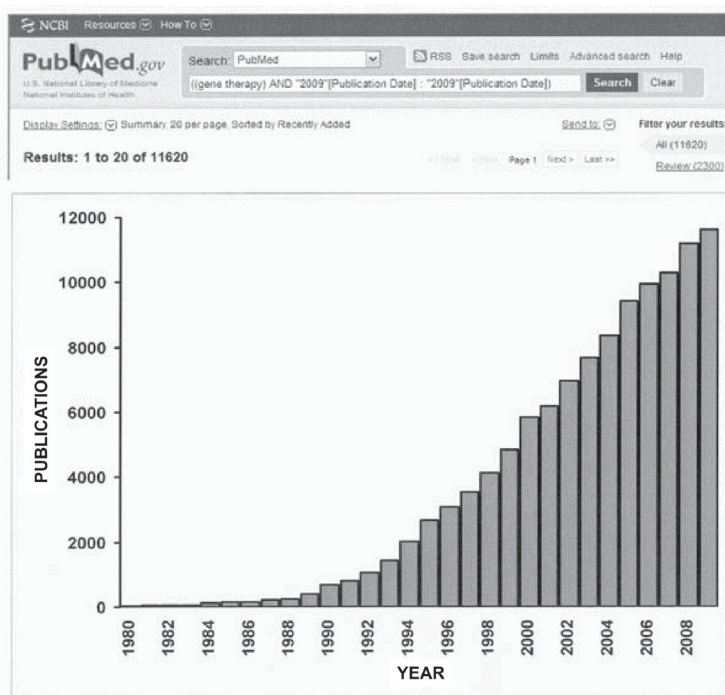


Figure 5 – Annual frequency of publication of scientific articles classified under the significant term “gene therapy”, registered in the database of the National Center for Biotechnology Information (PubMed) in the period 1980-2009.

As in other areas of research into novel therapeutic methods, the use of a gene therapy product or process depends on a series of clinical trials, which are classified by phases. It starts with the so-called phase I, which is aimed to test the safety of the procedure and identify any adverse effects attributed to the novel product or method. Next come trial phases II, III and IV which, progressively and always monitored for adverse effects, are intended to test the efficacy of the novel product or method in growing samples of patients, often distributed into multiple research centers.

Conducting these gene therapy trials depends on previous approval by local and national ethics committees such as the National Committee of Ethics in Research (CONEP) in Brazil, or the Food and Drug Administration (FDA) in the United States. In the case of gene therapy, in Brazil there is also the National Biosafety Technical Commission (CTNBio), and a specific committee of the National Institute of Health (NIH), called RAC (Recombinant DNA Advisory Committee) in the United States, which are responsible for authorizing procedures involving recombinant DNA. However, unlike in the United States, there is no specific gene therapy regulation in Brazil, which urgently needs to be developed both to

prevent the inappropriate use of therapies and to control the production and import of raw materials from abroad. Presently, it is up to the health authorities to enforce standards established abroad to examine any license requests or to oversee clinical trials and inspect any gene therapy products in the country.

Worldwide, nearly 1,650 clinical gene therapy trials had been registered in the database of the Journal of Gene Medicine (<http://www.wiley.co.uk/genmed/clinical/>) until June 2010. Figures 6-11 illustrate the main aspects of the current state of clinical research in this area.

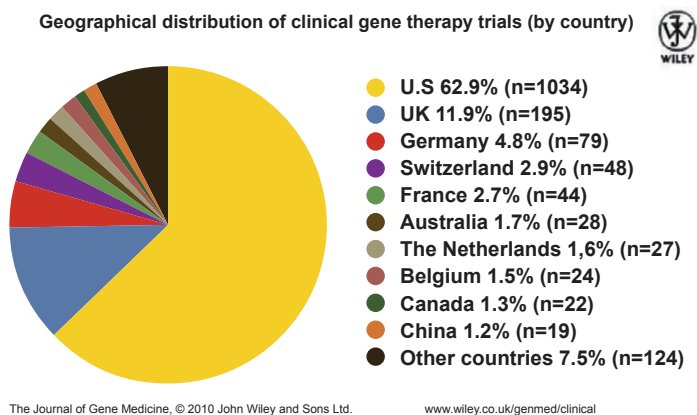


Figure 6 – Geographical distribution of countries hosting clinical gene therapy trials. Reproduced from the website of the Journal of Gene Medicine with permission of the publishers.

The distribution of countries hosting clinical trials (Figure 6) corresponds generally to the investment made in fundamental research in previous years. Among the countries that make up the “others” group, the JGM database includes one trial hosted in Mexico and none in South America. In fact, of the 38 clinical trials underway in South American countries identified at the end of 2009 on the database of the U.S. National Institute of Health (www.clinicaltrials.gov), 37 are extensions of trials hosted in countries of the northern hemisphere and only one, started in 2009, is actually hosted in South America, specifically in Brazil (see below).

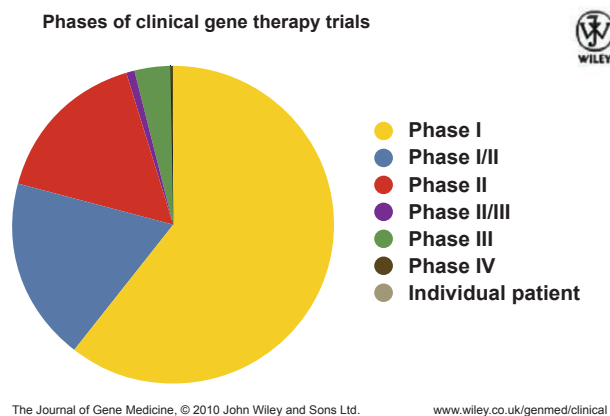


Figure 7 – Phases of clinical trials recorded in the database of the Journal of Gene Medicine. Reproduced with permission of the publishers.

The distribution into phases (Figure 7) clearly reflects the experimental character of gene therapy. For comparison purposes, data on the set of clinical trials registered on the webpage clinicaltrials.gov can be mentioned. Among these trials, which include mainly pharmaceuticals and conventional medical and surgical procedures, about 45 percent are phase II trials and just over 30 percent are phase III trials. In turn, as shown in the graph on Figure 7, the majority of clinical gene therapy trials are still in phase I and, to date, only about 4 percent have reached phases III and IV. However, there are signs that the progression of experimental gene therapy towards medical practice is gaining speed (Figure 8).

Safety is still the main barrier to the development of gene therapy into medical practice. The main obstacle is the fact that the safer non-viral vectors currently available are still little efficient or have very limited application, as is the case of the plasmids discussed above. The high efficiency of viral vector transduction makes the latter the most promising for application. However, some types, especially of adenoviral and retroviral vectors, which are the most widely used to date, have produced adverse effects, some serious and even fatal, and contributed greatly to the interruption of many studies in Phase I.

Of course, 150 years of fundamental research into pharmacology provide a solid foundation on which safety issues regarding conventional medicinal drugs are often solved in basic research laboratories or in well-established and highly predictive pre-clinical trials. There is still a long path to be trodden before this situation becomes routine in gene therapy research.

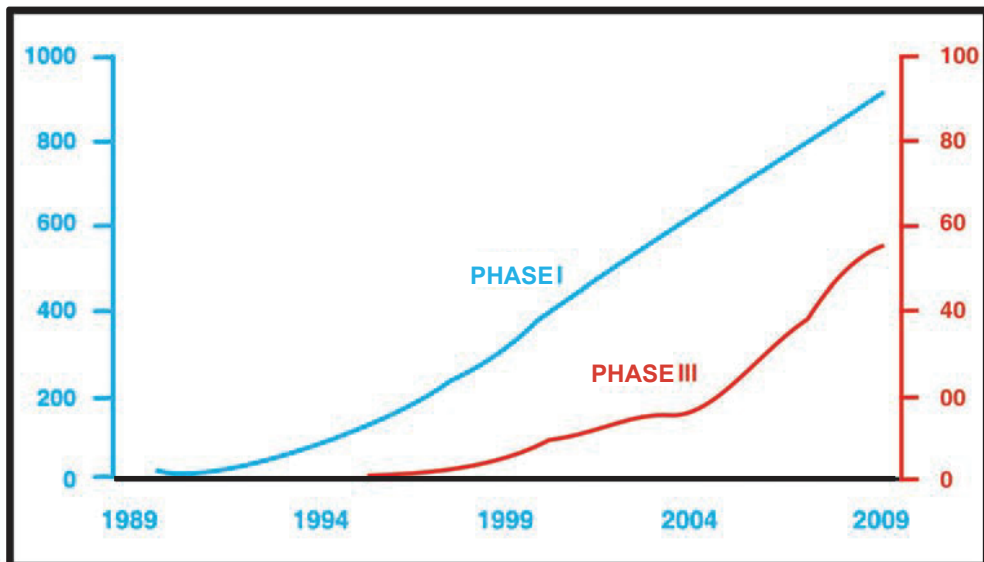
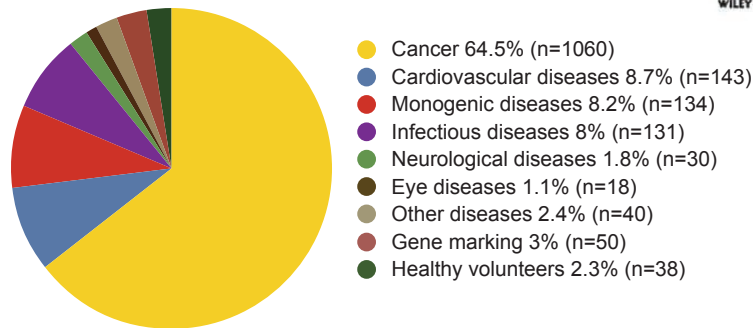


Figure 8 – Cumulative curves of evolution of phase I and phase III clinical trials in the area of gene therapy, developed from data registered in the database of the Journal of Gene Medicine. While phase I trials show a linear growth, the red curve suggests an acceleration in the evolution of phase III trials since 2004. Note that the vertical scales for the two phases are different.

Clinical Indications of Clinical Gene Therapy Trials



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www.wiley.co.uk/genmed/clinical

Figure 9 – Therapeutic indications of clinical trials registered in the database of the Journal of Gene Medicine. Reproduced with permission of the publishers.

The distribution of clinical trials by therapeutic indication (Figure 9) confirms a point previously raised in this article. Although gene therapy was originally conceived with the aim of treating monogenic diseases, these today account for less than 10 percent of clinical trials. The prevalence of cancer may be explained in part by greater ease in the approval of clinical trials based on the

compassionate use of experimental drugs or therapies in terminally ill patients, but also by the breakthrough in the design of oncolytic viruses (which destroy tumor cells) and suicide gene therapies (see below).

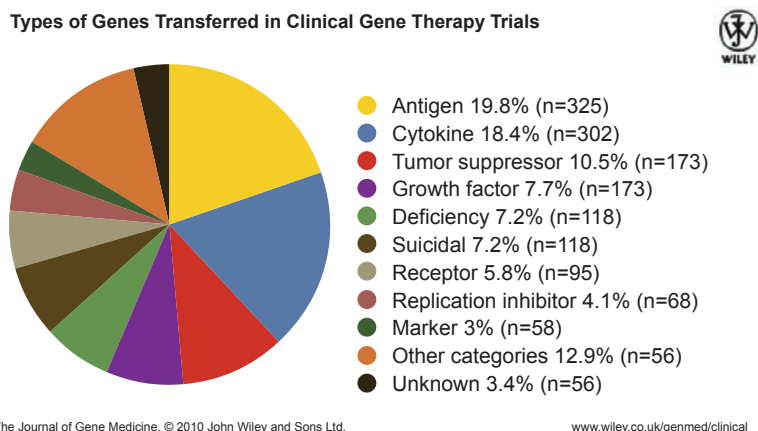


Figure 10 – Classification of genes used in clinical trials registered in the Journal of Gene Medicine. Reproduced with permission of the publishers.

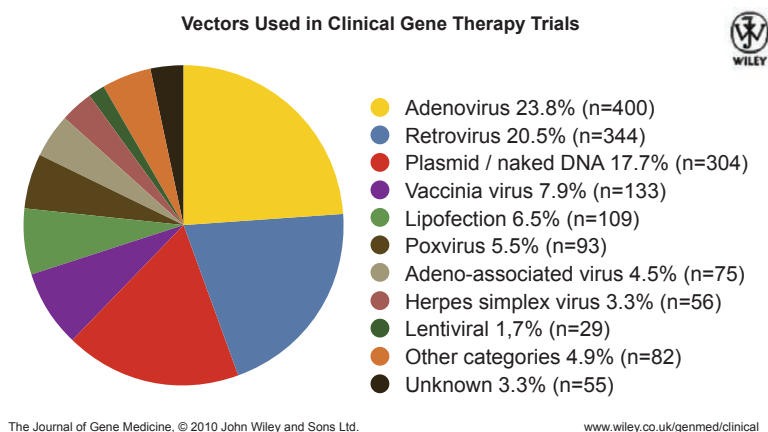


Figure 11 – Vectors used in clinical trials registered in the database of the Journal of Gene Medicine. Reproduced with permission of the publishers.

The variety of genes used in clinical trials (Figure 10) reflects the ad hoc nature of gene therapy. Progress in this area of medical research is probably strongly influenced by the trend towards the development of personalized medicine based on advances in genetics, pharmacogenomics and other fields of modern research. Still, the prevalence of antigens, cytokines, tumor suppressor genes and suicidal genes corresponds to the prevalence of cancer as the most frequent indication in clinical research in this area. The topic of vectors is undoubtedly the most critical to the advancement of gene therapy as regards application to medical practice. The graph in Figure 11 compiles data obtained

over two decades, during which technological progress in the field of vectors was extraordinary. For example, in contrast to the type of adenovirus vector that led to the death of a patient in a clinical trial in 1999 (see below) and almost paralyzed gene therapy research, third-generation adenoviral vectors are available today, which are radically modified in order to avoid adverse effects such as the one that killed the said patient. However, there is increased expectation in the use of viral vectors inherently safer, such as vectors derived from adeno-associated viruses.

Applications of gene therapy

To illustrate the potential applications of gene therapy as well as the underlying logic and the sequence of fundamental and pre-clinical research that led to clinical trials, some examples have been selected, which are described below.

Monogenic diseases

Haemophilia: Since each type of hemophilia is a monogenic disease, the procedure is to introduce the respective healthy gene (Factor VIII or Factor IX, depending on the type of hemophilia) into the patient's cells, so that these will start to produce the required protein. The therapy should not only get the body to produce the missing protein, but to produce it in sufficient quantity to restore the patient's health and for a long time, ideally throughout the person's lifetime.

After extensive preclinical trials in mice and dogs, which showed long-term recovery of factor IX infusion-mediated procoagulant activity using experimental gene therapy trial, two Phase I / II studies have been recently conducted by a group at the University of Pennsylvania, with the infusion of an adeno-associated virus vector (rAAV) containing the gene encoding factor IX of haemophilia B patients (Manno et al., 2003, 2006, Hasbrouck & High, 2008). No serious adverse effects were observed in any of the patients tested.

The results indicate the potential efficacy of the treatment, since a patient who received a high dose of rAAV-F9 by liver infusion presented, between two and five weeks of treatment, therapeutic levels of circulating factor IX above 10 percent of normal activity, which is sufficient to sustain blood coagulation capacity. However, the therapeutic effect was transient, having disappeared six weeks after treatment, accompanied by a temporary and asymptomatic increase in transaminase levels (Figure 12). The results observed in this patient as well as in another patient in the same study indicated that the therapeutic effects disappeared as a result of degeneration of the liver cells in which the vector had been introduced, caused by an immune response to the viral vector proteins (Mingozzi & High, 2007).

This example is particularly important because neither in any of the pre-clinical trials conducted in animals prior to the clinical study, nor in new experiments conducted after the results of the said clinical trial were achieved, immune responses were observed in the animals tested that would enable predicting the

immune response observed in the patients. The result demonstrated the need for caution in the transition from preclinical to clinical trials, even in the absence of serious adverse effects, and provided critical data to advance the clinical application of gene therapy. New experimental studies are underway, with the aim to avoid this immune response with the use of vector variants and transient immunosuppression, which will guide future clinical trials (Hasbrouck & High, 2008).

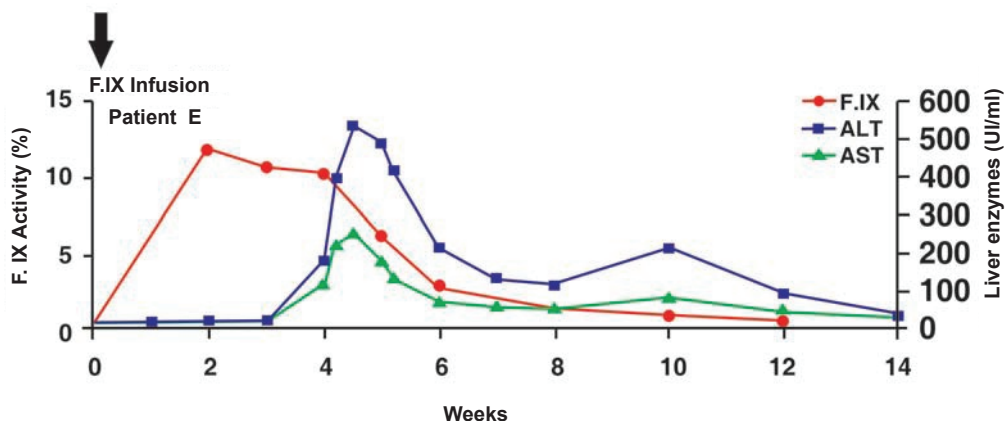


Figure 12 – Activity of circulating factor IX (red) and transaminase levels (blue and green) over time after gene therapy for hemophilia B in a clinical trial conducted with the use of adeno-associated virus vector containing the gene encoding factor IX. Modified from Hasbrouck & High (2008).

Leber’s congenital amaurosis: The first results of phase I/II clinical trials for the treatment of Leber’s congenital amaurosis (LCA) began to be published in the end of April 2008. LCA is a disease that causes progressive blindness, starting with significant loss of vision in infants and progressing over time to total blindness. Initially, photoreceptors, which are light-sensitive retinal cells essential for vision, are inactivated, but remain alive in the retina (den Hollander et al., 2008). Over the years, inactive photoreceptors, predominantly rods that function in dim light, degenerate and disappear (Spuy et al., 2005).

There are several forms of LCA, some of well-known genetic cause, such as RPE65 deficiency, an enzyme required to produce the vitamin derivative required for the functioning of photoreceptors (Poehner et al. 2000; Bereta et al. 2008). The photoreceptors of these patients gradually lose function, but degeneration usually only occurs around 30 years of age (Hollander et al., 2008). The course of the disease provides a therapeutic window for injecting healthy copies of the RPE65 gene into the retina of young adults with this form of LCA (Figure 13). The tests are still preliminary and, in principle, only three patients have been tested in each of the three Phase I clinical trials conducted in England and in the United States (Bainbridge et al., 2008; Maguire et al., 2008; Cideciyan et al., 2008; Hauswirth et al., 2008).

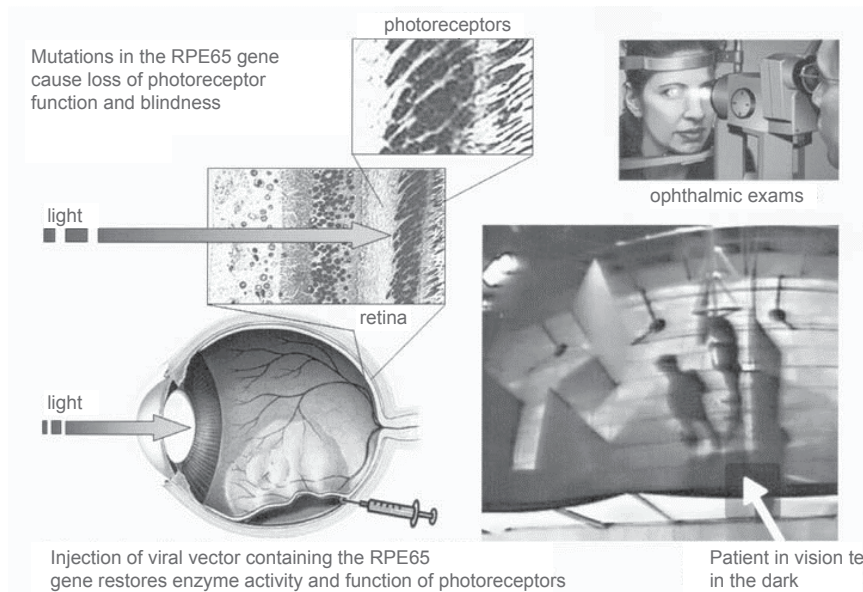


Figure 13 – Diagram of gene therapy for Leber congenital amaurosis caused by RPE65 mutations. The figure illustrates clinical trials conducted since 2007 by teams from the University College in London, England, and the universities of Pennsylvania and Florida, in the United States. This degenerative retinopathy initially produces inactivation of rod function, with progressive loss of scotopic vision and subsequent photoreceptor degeneration before 30 years of age, leading to blindness. The three research groups are testing the effects of the injection of healthy RPE65 gene into the retina of young adult patients with LCA. Studies are underway and researchers are evaluating also the safety of the procedure, the patients' visual acuity compared to that presented before the gene therapy. In some cases there was an improvement in the results of ophthalmic exam (top right), pupillary reflexes and spatial orientation in dimly lit environments containing various obstacles (below right, in a frame extracted from the movie *Moorfields Hospital*, courtesy of Prof. Robin Ali, University College, London).

The first results showed that the infusion of adenovirus-associated virus vectors containing the healthy gene in the patients' retina did not cause significant adverse effects. Improvement was observed in ophthalmic exams and visual performance in some patients, who partially recovered sensitivity to light (Hauswirth et al., 2008) and the ability to orient themselves in dimly lit environments, something they were unable to do before the injection of the healthy gene (Bainbridge et al., 2008).

The results achieved so far, however, relate to few patients, and no signs of improvement have been observed in certain crucial ophthalmic exams (Hauswirth et al., 2008), and RPE65 deficiency accounts for only 6 percent of LCA cases (Hollander et al., 2008). That is, the therapy that is being tested at the moment, if successful, can only be applied to a small fraction of patients. Treat-

ments for the other groups of patients need to be developed on a case by case basis. Still, it is an important advance in the development of novel therapies for blindness-causing diseases, and a phase II clinical trial is underway to confirm (or not) in a systematic way, the possible efficacy of the treatment.

Cancer

Most clinical gene therapy trials have been conducted in cancer patients (Figure 9), usually in advanced stages of the disease. The desirable effect of any cancer treatment is to cause the selective death of tumor cells (Evan & Littlewood, 1998; Green & Evan, 2002). Cancer cells often multiply rapidly, which explains the growth of tumors. Many drugs are used in treating cancer precisely because they selectively attack cells that multiply rapidly and, therefore, kill tumor cells (Wang et al. 2,008; Prochownik, 2008; Vazquez et al. 2008).

The physiological need for continuous renewal of blood cells from the proliferation of bone marrow precursors implies, however, severe adverse effects of chemotherapy. These effects are difficult to avoid because, among other factors, the drugs are injected into the bloodstream. For treating cancer it is desirable to reach, in some way, only the tumor cells. In the case of solid tumors, such as tumors in the central nervous system, this can be achieved through localized gene therapy (Rainov & Ren, 2003), and various strategies are being developed accordingly (Table 2).

The procedure dubbed “suicide gene technique” consists in introducing into the tumor cells a gene that does not exist in the human genome and that encodes the thymidine kinase enzyme of the herpesvirus genome. The presence of this enzyme in a human cell kills the cell in the presence of a drug called ganciclovir, as the thymidine kinase converts ganciclovir into a toxin. The toxin, in turn, affects only multiplying cells (Figure 14).

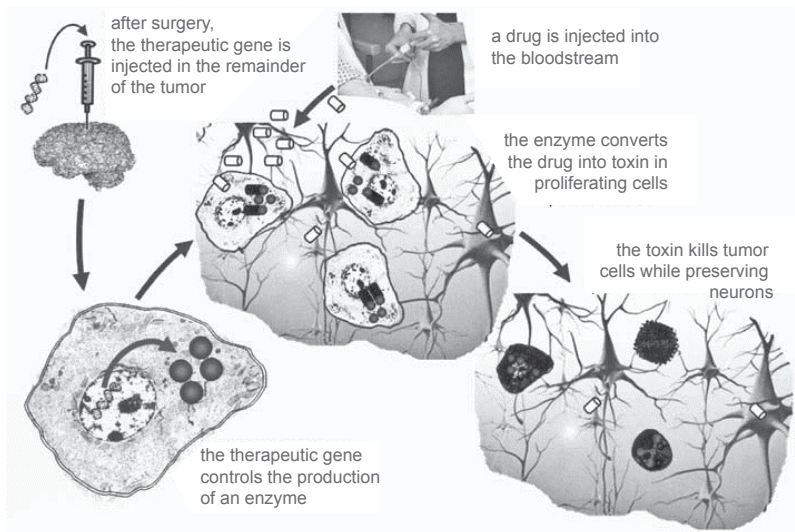


Figure 14 – Action mechanism of suicide genes. The concept, formulated in the late 1960s by the American researcher Frederick Moolten from Boston University, is

shown schematically in the figure, following the arrows from the upper left corner. The example refers to glioblastomas, but also applies to other types of tumors. First, the neurosurgeon removes as much of the tumor as possible, leaving tumor cells scattered among normal brain neurons. The therapeutic gene (HSV-TK or thymidine kinase) is injected into the surgical area, penetrating the cells and commanding the production of the enzyme. This enzyme phosphorylates the ganciclovir injected, transforming it into a powerful toxin that is incorporated into the DNA of target cells, blocking DNA replication and leading eventually to the death of the proliferating cell. Reproduced from Linden (2008) with permission of the publishers.

Although the efficacy of suicide gene technology to treat tumors is still controversial, some studies have reported promising results. Among them is a phase I/II clinical trial conducted in Finland, in which the resection of extremely aggressive tumors of the central nervous system, known as glioblastomas, was followed by the injection into the surgical cavity of an adenoviral vector containing the thymidine kinase gene of herpesvirus. The procedure continued with daily intravenous injections of ganciclovir for 14 days. Gene therapy resulted in a significant increase in the survival (Figure 15) of the group of 17 patients treated with gene therapy, as compared to a group of 19 patients treated with conventional therapy, or as compared to a control population of 36 patients previously treated by conventional methods in the same neurosurgery unit, in the two years prior to the trial (Immonen et al., 2004). The vector used in this study is being developed by the company Ark Therapeutics, which recently reported significant positive results of a phase III multicentric study with 250 patients, and in February 2009 was granted in France the first authorization for compassionate use of the product called Cerepro®.

Table 2 – Genes and gene therapy strategies for tumors of the central nervous system

Strategy	Examples	Operation
Suicide genes - induction of programmed selective killing of tumor cells	HSV-TK (herpes virus thymidine kinase)	Blocking of DNA synthesis in the presence of a pro-drug
Oncolytic virus with conditional replication	HSV-1 Onyx-015	Replication only in dividing or tumor cells
Induction of apoptosis	FasL, TRAIL	Activation of apoptosis
High affinity ligands	Transferrin receptor	Drug specifically targeted to the tumor
Corrective strategy	p53, Rb, p16, PTEN	Correction of genes eliminated from tumors
Immune gene therapy	Interleukins, interferons, TNF- α	Activation of antitumor immune response
Suppression of angiogenesis	Angiostatin, endostatin	Blocking of blood vessel growth
Interference RNA	VEGF, EGFR, IGFR	Decreased expression of oncogenes
Combination with cell therapy	Neural stem cells or mesenchymal as producers of viral vectors	Continuous and localized production of viral vectors

Source: Adapted from Linden & Lenz (2007).

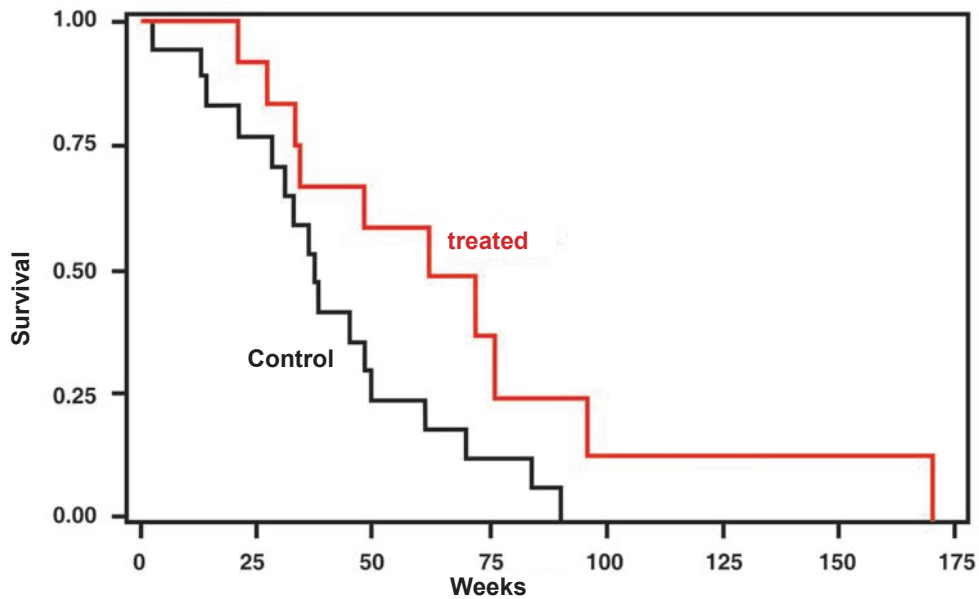


Figure 15 – Increased survival of glioblastoma patients treated with gene therapy with the use of “suicide gene”. An adenoviral vector encoding the gene of thymidine kinase enzyme was injected into the surgical cavity after resection of the tumors, followed by intravenous injections of ganciclovir for two weeks. The Kaplan-Meier curve indicates the fraction of surviving patients over time for patients treated (red) and controls (black). The interpretation is that gene therapy was effective on part of the tumor cells that remained after surgery, which could not be attacked otherwise. Modified from Immonen et al. (2004).

Parkinson’s Disease

Neurodegenerative diseases are one of the most complex classes of diseases facing contemporary medicine. Despite the advances achieved since the 1990s, a period called “the brain decade” (Goldstein, 1994), and the extensive body of knowledge about various aspects of the pathogenesis, genetics, clinical course, complications and response to different treatments tested over years of research, there is a conspicuous lack of therapeutic options, particularly in the later stages of these diseases (Radunovic et al., 2007; Cacabelos, 2007; Han & McDonald, 2008; Jalbert et al., 2008; Gauthier & Poirier, 2008; Olanow et al., 2008).

In turn, some neurodegenerative disorders illustrate the potential for the development of gene therapy for multifactorial and high complexity diseases. Parkinson’s disease (PD) is an example of this category.

PD is characterized by the progressive death of neurons in the substantia nigra pars compacta of the midbrain and functional changes in other brainstem nuclei (Figure 16), followed by the appearance of intracellular inclusions known as Lewy bodies. This results in loss of dopamine - the neurotransmitter used by neurons that degenerate - in the target of nigral extension neurons in the substantia nigra, which is called the striatum. As the disease progresses, other

neurotransmitter systems become involved. Motor disorders typical of the disease, such as resting tremor, slowness of movement and muscle stiffness are frequently accompanied by postural instability, visceral dysfunction and cognitive disturbances (Guttman et al., 2003). The mechanisms that lead to the death of nigral neurons are still controversial (Dawson & Dawson, 2003; Dauer & Przedborski, 2003).

Pharmacological treatment with L-dopa, a precursor of dopamine synthesis, is effective in the short or medium term, but tends to become innocuous with the progressive loss of neurons, and can potentially cause additional motor disorders. The progression of the disease requires higher doses and combinations of drugs, which not always are effective (Poewe, 2009). Cellular therapies aimed at regenerating nigral dopaminergic neurons may eventually benefit PD patients, but so far clinical trials conducted with fetal nerve cell grafts have produced discrete effects and suggested the possibility of disease transmission to the transplanted tissue (Thajeb et al. 1,997, Li et al. 2,008; Kordower et al. 2,008; Mendez et al. 2,008; Braak & del Tredici, 2008).

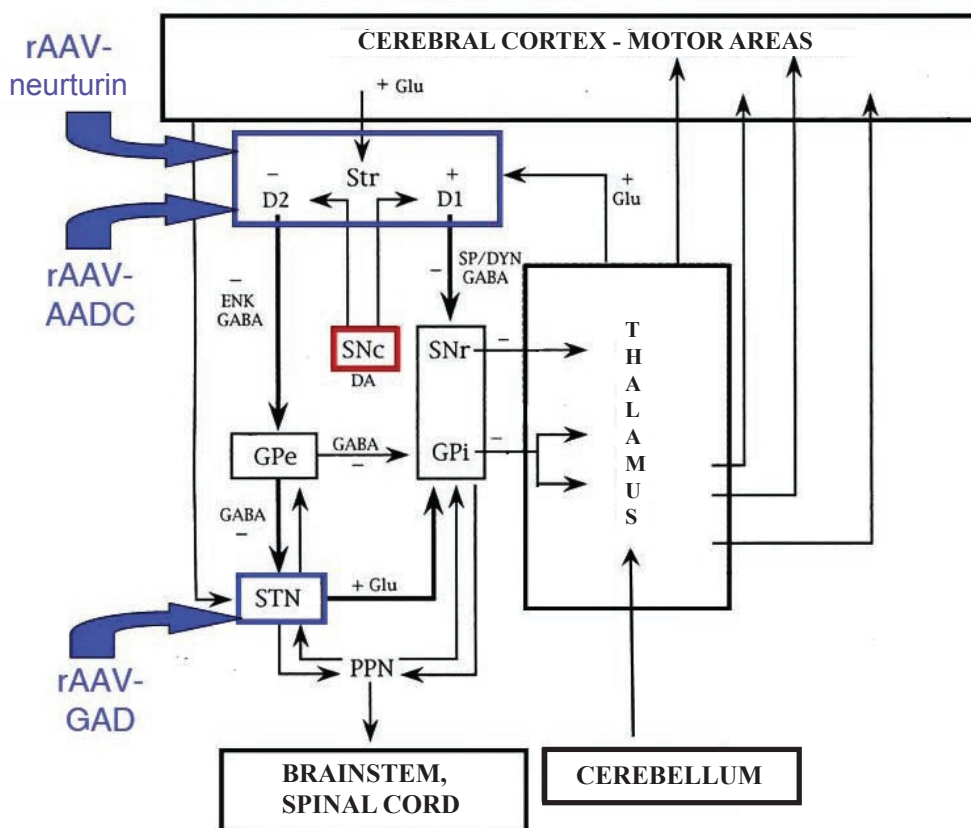


Figure 16 – Diagram of main neurotransmitter connections and systems of the basal ganglia circuitry relevant for Parkinson's disease. Degeneration (in red) of substantia nigra pars compacta dopaminergic neurons (SNc) reduces

activation of dopamine receptors (D1 and D2) in the striatum (Str). As a result, the activity of projection targets of the striatum becomes unbalanced producing, among other effects, hyperactivity of the glutamatergic neurons of the subthalamic nucleus (STN), causing motor disturbances. The strategies of gene therapy trials in PD patients are indicated by arrows and their respective targets are in blue. Modified from Nakano (2000).

Gene therapy strategies for Parkinson's disease include induction of local production of dopamine in the striatum, supply of neurotrophic factors to reduce the progressive loss of dopaminergic neurons, or even compensation of functional imbalance in the cell communication network of the basal ganglia (Chen et al. 2005).

The production of dopamine depends mainly on the activity of three enzymes. The techniques aimed to produce dopamine in the depleted striatum involve, in general, viral-vector induction of one or more of these enzymes (Kang et al. 2001). Preclinical experimental models consist of chemical lesions of the substantia nigra in rats or primates. Several types of viral vectors have been tested (Chen et al., 2005 for review). Based on the results of preclinical studies, a phase I clinical trial has been started to test the safety and, secondarily, the beneficial effects of gene therapy by expression of one of the producing enzyme (AADC), carried by an adenovirus-associated vector injected in the striatum of PD patients who have had the diseases for 14 years on average (<http://clinicaltrials.gov/show/nct00229736>). The results (Christine et al., 2009) showed clinical improvement without adverse effects of the gene therapy per se, although risks have been identified in the surgical procedure.

In turn, neuroprotection strategies aimed at reducing or preventing neuronal loss in the long term have been developed based on several growth factors that have a protective effect on nigral neurons. Experiments in animals include a study that tested the effects of the injection, in the striatum, of a viral vector expressing a neurturin gene construct, with positive results (Fjord-Larsen et al., 2005). Thus, a phase I clinical trial was initiated in mid-2005, with a view to examining the safety of an adeno-associated viral vector expressing neurturin gene injected into the striatum (<http://clinicaltrials.gov/show/nct00252850>). No serious adverse effects were observed in 12 patients treated with two different doses of the vector, and beneficial effects were detected in some motor parameters (Marks et al., 2008). A multicentric Phase II study is currently underway.

The third gene therapy strategy for PD is based on the functional imbalance between excitatory and inhibitory pathways in the basal ganglia, caused by the loss of substantia nigra activity (Figure 16). Under these conditions, there is disinhibition of the activity of a part of the brain called the subthalamic nucleus (STN), to which an important role in the main signs of PD is attributed (Nakano, 2000; Chen et al. 2005). Several studies have shown that surgical removal of the STN or high frequency electrical stimulation has beneficial effects

on some of these signs, justifying the use of the so-called deep brain stimulation in the treatment of advanced cases of PD (Jankovic & Diamond, 2005). Knowledge of the functional properties of neural circuits involved in the disease has led to a striking example of genetic intervention designed to modulate the physiology of the nervous system, regardless of the cause of the disease which, even today, remains controversial.

A gene therapy trial has been developed, which involves inducing the expression of enzymes that produce an inhibitory neurotransmitter, with the aim of inhibiting excessive neural activity in the STN. The expression of these enzymes in the STN produced beneficial functional effects in PD models in rats (Luo et al. 2002). Based on these results, a phase I clinical gene therapy trial was conducted in the period 2003-2005, using a recombinant adeno-associated virus vector containing the gene encoding one of these enzymes, injected into the STN (<http://www.clinicaltrials.gov/ct/show/nct00195143>). Results for 11 patients monitored for up to 12 months showed significant improvement in motor performance, accompanied by reduced metabolic activity in STN projection targets, consistent with the results of preclinical studies. A significant improvement in daily activities was also reported, which reflects the views of patients about their performance on tasks of everyday life. No adverse effects have been reported questioning the safety of the procedure (Kaplitt et al., 2007).

The results of the clinical trials described are still very preliminary, were obtained in small numbers of patients and require confirmation through broader trials with more stringent controls for placebo effects and other variables. Therefore, it is still early to conclude on the viability and particularly the efficacy of gene therapy for neurodegenerative diseases. However, these studies are in addition to other clinical trials suggesting that gene therapy may become an effective alternative for treating currently incurable diseases.

Risk-benefit assessment of gene therapy

Among the hundreds of clinical gene therapy trials completed to date, most aimed to test the safety of the procedure. In certain cases, early detection of adverse effects during the study period was sufficient to immediately terminate the trial, thereby avoiding any risk of aggravation. But in many cases, the procedure used was considered safe, with only occasional, mild and tolerable adverse effects.

Mild pain or inflammation at the injection site, mild transient fever, temporary headache, flu-like symptoms and other mild effects are generally tolerable in view of the potential for treatment of an incurable disease. These are most of the incidents that are usually found in phase I clinical gene therapy trials, especially after the completion of extensive preclinical tests in animals, as required by regulatory agencies for authorizing clinical trials (information on regulatory agencies in the U.S. and Europe and on regulation in this area can be found, for example, at <http://www.genetherapynet.com/legislation.html>).

Immune reactions, however, not only can cause adverse effects, but even in the absence of these can destroy the vectors or cells infected with viral vectors, despite the use of sophisticated recombinant DNA techniques in their production. This was the case of the clinical trial for hemophilia B previously described (Mingozzi & High, 2007), but which did not bring significant consequences for patients. In other cases, however, the adverse effects can be quite severe or, in rare cases, even fatal.

In 1999 a patient died soon after the injection of a viral vector during a clinical gene therapy trial, victimized by a systemic inflammatory response syndrome caused by first-generation adenoviral vector (Raper et al., 1998, 2003). In more recent clinical trials conducted in France and England (Hacein-Bey-Abina et al., 2002; Gaspar et al., 2004), of a total of 20 children under one year of age undergoing gene therapy for X-linked severe combined immunodeficiency syndrome (XL-SCID) (Buckley, 2004), five developed leukemia (Hacein-Bey-Abina et al., 2003; Howe et al., 2008). Of these, one died and four went into complete remission after chemotherapy. Tests performed after the onset of leukemia revealed that the retroviral vectors used in both trials produced insertional mutagenesis, i.e., mutations produced by the injection of the DNA vector, breaking the continuity of the genetic sequence (Cavazzana-Calvo & Fischer, 2007; Howe et al., 2008).

The above mentioned cases are the most serious examples actually characterized as direct adverse effects of gene therapy. Both are derived from characteristics of the viral vectors used. However, in both cases fundamental research coupled with careful observation of the events associated with the treatment and the clinical course of the adverse effects contributed to advances in the design and production of novel vectors aimed to avoid such adverse effects.

In the case of adenoviral vectors, in contrast to the first-generation vectors used in the clinical trial that resulted in the fatal case in 1999, third-generation adenoviral vectors constructed with complete deletion of viral genes and capable of much safer gene transduction in humans are now available (Räty et al., 2008; Dormond et al., 2009). In turn, there is a growing expectation of avoiding insertional mutagenesis like the one observed in XL-SCID trials, through the design of auto-inactivating retroviral or lentiviral vectors or vectors with chromatin insulators, two of the most promising techniques currently under development for this vector class (Yi et al., 2005; Räty et al., 2008).

Gene therapy for XL-SCID, in turn, was curative in 19 of the 20 children treated, who showed significant improvement in their immune system less than three months after treatment, as well as persistent recovery of resistance to infections (Table 3; Fisher & Cavazzana-Calvo, 2008; Aiuti Roncarolo, 2009). However, the treatment was not effective in adolescents, suggesting a limited therapeutic window for intervention in this disease. Adding up to successful cases are 30 patients treated for ADA-SCID, the form of immunodeficiency that corresponds to the first patient treated by gene therapy in 1989 (Table 4; Aiuti & Roncarolo, 2009).

Regulatory agencies involved in the authorization and control of clinical

gene therapy trials acted quickly in both cases of adverse effects reported here. In 1999, the trial that resulted in the patient's death was permanently discontinued, despite the absence of serious adverse effects in the other 17 patients treated in the same study. In the case of XL- SCID trials, therapeutic procedures had already been terminated, but authorizations for other similar trials were suspended pending a thorough evaluation of the data, and were subsequently granted. Despite the recognition that the gene therapy procedure was responsible for the adverse effects, the regulatory committees concluded that none of these events, as well as other adverse effects reported occasionally justified the suspending clinical gene therapy trials. Indeed, the analysis of adverse effects has helped guide the development of biotechnology in the field and, at the same time, improve regulation and criteria for clinical trial authorization.

Table 3 – XL-SCID clinical gene therapy trials

Trial	No. of patients	Observation period	Effectiveness	Toxicity
Necker Hospital, Paris	10 (age < 1 year)	10 years	yes	Leukemia (4 patients), 3 complete remissions after chemotherapy
Great Ormond St Hospital, London	10 (age < 1 year)	7 years	yes	Leukemia (1 patient), complete remission after chemotherapy
Multicentric, FR, UK, US	5 (age = 10-20 years)	3 years	no	no

Source: Adapted from Aiuti & Roncarolo (2009).

Table 4 – ADA-SCID clinical gene therapy trials

Trial	No. of patients	Observation period	Effectiveness	Toxicity
HSR-TIGET	15	8 years	yes	no
GOSH	5	5.5 years	yes	no
CHLA/NIH (1)*	4	8 years	no	no
CHLA/NIH (2)*	6	2 years	yes	pancytopenia** (1 patient)

* Studies (1) and (2) differ in the pre-drug treatment of patients before gene therapy.

** The adverse effect was attributed to a cytogenetic abnormality independent of the gene therapy.

Source: Adapted from Aiuti & Roncarolo (2009).

The assessment of benefits and adverse effects in clinical gene therapy trials indicates that the course of their development, as well as of other advanced therapies such as stem cell treatments will be much safer when well substantiated by basic research and subject to appropriate regulations to condition the authorization of clinical trials upon maximum safety condition at the time of trials, but without inhibiting the advancement of medical research.

Gene therapy and biotechnology

Entrepreneurs in the area of biotechnology saw in human genome sequencing growing business opportunities. The interest lies, of course, in the fact that the discovery of genes and especially of mutations responsible in whole or in part for a disease can lead to the development of diagnostic tests or marketable drugs.

Among other things, companies have begun to invest in the patenting of genes or sequences of DNA fragments that had not even been associated with genes. More than three million genome-related patents have been filed to date in the United States. U.S. legislation, in general, authorizes the patenting of genes, provided that these are isolated (and not just described as nucleotide sequences) and supported by evidence of usefulness, for example, for the development of diagnostic tests. However, the patenting of genes is controversial. For example, the internal rules for evaluating the usefulness of findings related to genes enforced since 2001 by the Office of the United States Patent Office (USPTO) have been and still are the subject of severe criticism, from which the USPTO defended itself on the grounds of the patent legislation in force in the United States. On the other hand, the National Institute of Industrial Property (INPI), the Brazilian agency that grants nationally valid patents, reports on its website that the patenting of natural genes is prohibited in Brazil.

Outside the scope of the controversy over the patenting of genes, vectors for gene therapy, whether viral or nonviral, containing therapeutic genes, as well as their specific applications are products of technological development and, as such, are legitimate objects of patenting and possible marketing (Bobrow & Thomas, 2002). Hundreds of such patents have been filed with the USPTO and its counterparts in Europe and Asia. Dozens of companies are investing in gene therapy, based on patented technologies for the production of vectors or as partners of research institutions (Table 5).

Worldwide, the first development stages of gene therapy technologies and many preclinical trials are available to research groups, institutes and public universities as well as to private entities financed with public funds. However, the transfer of laboratory research to clinical trial usually requires resources that are far beyond the capacity of public funding. Biotechnology companies invest in such tests because of the existence of patents that may eventually be commercially exploited, as in all other areas of technology. To date, only one product specifically classified as gene therapy has been marketed, but other four products are at an advanced stage on the path to marketing (Table 6).

Table 5 – Overseas companies engaged in gene therapy

1. Advanced Cell & Gene Therapy, LLC	28. Collateral Therapeutics, Inc.	58. Nephros Therapeutics, Inc.
2. Advanced Cell Technology	29. Copernicus Therapeutics Inc	59. Neurotech SA
3. Advanced Vision Therapies,	30. Corgentec Inc.	60. Nucleonics Inc.
4. AlphaVax Human Vaccines, InC.	31. Cyclacel Limited	61. Oncosis
5. Altogen Biosystems	32. CyThera Inc.	62. Onyx Pharmaceuticals, Inc.
6. Amaxa GmbH	33. Cyto Pulse Sciences, Inc	63. OrphagenicX
7. Amsterdam Molecular Therapeutics	34. CytoGenix Inc	64. Oxford BioMedica
8. Applied Tissue Technologies LLC	35. DeveloGen AG	65. PharmaFrontiers Corp.
9. Ark Therapeutics Ltd.	36. Enzo Biochem, Inc.	66. Polygenetics Inc.
10. Athersys, Inc.	37. Epeius Biotechnologies Corporation	67. PrimeGen Biotech LLC.
11. AuRx Inc	38. Expression Genetics Inc	68. Progenitor Cell Therapy, LLc
12. Austrianova FSG {FSG AUSTRIANOVA GmbH}	39. geneRx+	69. Proneuron Biotechnologies, Inc.
13. Avaris AB	40. Genetix Pharmaceuticals Inc	70. Regulon Inc.
14. Avigen Inc.	41. GenVec. Inc.	71. Reneuron
15. Bavarian Nordic A/S	42. IC-Vec Ltd	72. REPLICor Inc.
16. BetaStem Therapeutics Inc	43. Ichor Medical Systems, Inc.	73. Ribozyme Pharmaceuticals Inc. (RPI)
17. BioCardia Inc	44. Immuno-Designed Molecules (IDM), SA	74. Sertoli Technologies Inc.
18. Bioheart, Inc.	45. Insert Therapeutics Inc.	75. Stem Cell Sciences
19. BioProtein I Technologies	46. Intercytex Limited	76. StemCells Incorporated
20. Biovex Limited	47. Introgen Therapeutics, Inc.	77. Supratek Pharma Inc.
21. Cardion AG	48. Intronn, Inc.	78. Targeted Genetics Corporation
22. Cell Genesys Inc	49. Invivogen	79. TheraCyte, Inc.
23. Collectis SA	50. Ixion Biotechnology	80. Theratechnologies
24. Cellerant Therapeutics Inc.	51. MaxCyte, Inc.	81. TheraVitae Co., Ltd.
25. CellGenix Technology Transfer GmbH	52. MediGene, Inc.	82. Tosk Inc
26. Cellprep S. A.	53. Mirus Corporation	83. Transgene S.A
27. Ceregene Inc.	54. Innovata pic	84. Valentis, Inc.
	55. Molecular Medicine, LLC	85. VIA Pharmaceuticals, Inc.
	56. Mologen Holding AG	86. Vical Incorporated
	57. Nature Technology Corporation	87. VirRx, Inc.

Source: Data from International Scientific Products Exchange, 2009. Available at: <<http://www.ispex.ca/companies/genetherapy.html>>.

Table 6 – Gene therapy products in advanced stages of development

Product	Composition	Indication	Company	Status
Gendicine®	rAd-p53	Tumors of the head and neck	SiBiono GenTech, China	Approved and on the market in China (2003)
Rexin-G®	tumor matrix (collagen)-targeted, retroV-dnG1-Cyclin	Solid tumors	Epeius Biotech, U.S.	Compassionate use in Japan (2007); approved in the Philippines
Collatogene®	Plasmid-HGF	Critical limb ischemia	AnGes MG/ Daiichi Sankyo, Japan	Under review in Japan; special protocol analysis (SPA) in the U.S.
Advexin®	rAd5CMV-p53	Tumors of the head and neck	Introgen, U.S.	Under review; phase II Trial underway
Cerepro®	rAd5-TK	Glioblastoma	Ark Technologies, ING/Finland	Approved for individualized use in France and Finland (2009)

The interest of the industrial sector in gene therapy can be illustrated by data from organizations specializing in technological prospecting. The analysis of the evolution in the number of gene therapy products under development by companies reveals an important aspect. While scientific production in the area grows continuously (Figure 5), the growth curve of industrial investment showed a clear downfall between 2003 and 2007 (Figure 17), probably influenced by the adverse effects of XL-SCID gene therapy trials, which were widely disseminated and, of course, must have aroused concern among investors. Still, the number of industrial products in phases II and III continued to grow during the period (Figure 18) and the recovery of the sector's growth is predictable. In the business world there is a growing expectation of success at a timescale compatible with investments both in academic research and in the private sector (Phacilitate, 2009). A strategic study conducted in 2008 predicted a world market for gene therapy products of about \$500 million by 2015 (Global Industry Analysts - Gene Therapy: a global strategic business report, 2008).

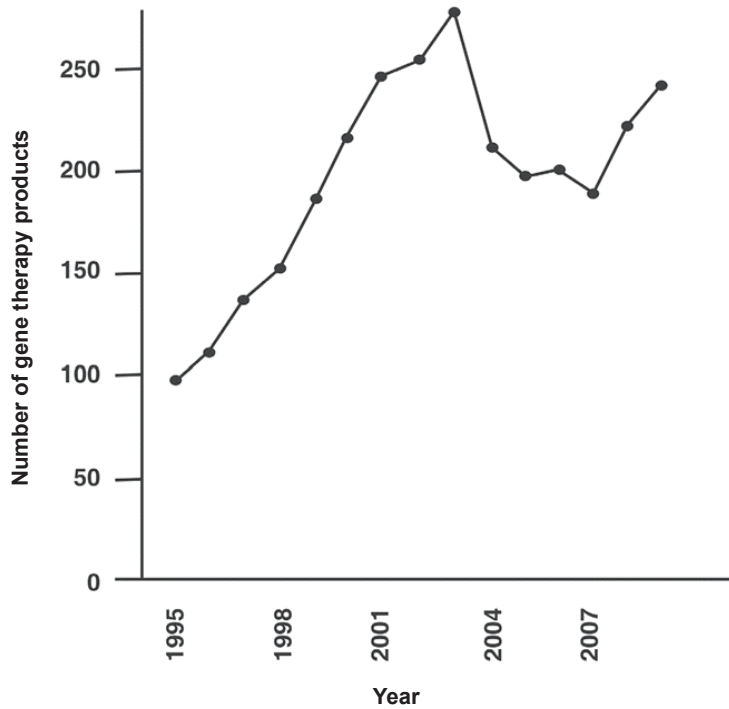


Figure 17 – Evolution in the number of gene therapy products under development in biotechnology companies in the period 1995-2009. Modified from <http://www.pharmaprojects.com/therapy_analysis/genether_early_0409.htm>.

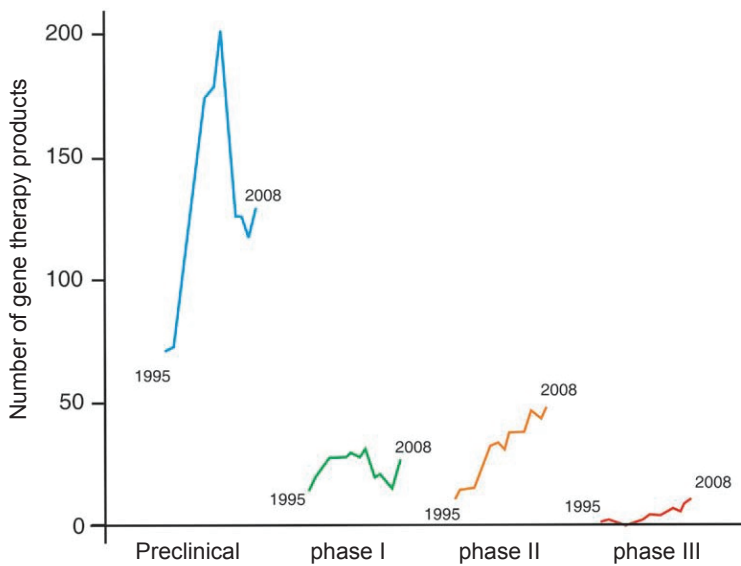


Figure 18 – Evolution in the number of gene therapy products in preclinical or clinical trial phases under development in biotechnology companies in the period 1995-2008. Modified from http://www.pharmaprojects.com/therapy_analysis/genether_current_0409.htm.

Gene therapy in Brazil

Despite the history and international recognition of Brazilian genetics, there are few research groups devoted to gene therapy studies, including DNA vaccines. Until recently there was little public investment in this research area and no interest from the private sector. The picture, however, is starting to change with some initiatives, albeit modest, in both sectors.

The Gene Therapy Network

A Gene Therapy Network began to be organized in Brazil in 2005. This network, coordinated by this author, initially brought together 14 research groups from three states (Rio de Janeiro, São Paulo and Rio Grande do Sul), dedicated to research in the area of gene therapy and DNA vaccines. The studies involve the development of viral vectors, basic research and preclinical trials in the areas of cancer, genetic diseases, neurodegenerative diseases and DNA vaccines for dengue fever, Chagas disease, streptococcal infections and cancer.

A first clinical gene therapy trial for myocardial revascularization with the use of plasmid vectors containing the VEGF gene (Vascular Endothelial Growth Factor), was started in February 2009 in Porto Alegre, promoted jointly by the Institute of Cardiology of Rio Grande do Sul, the Research Support Foundation of Rio Grande do Sul and the Gene Therapy Network, through the MCT/CnPq Millennium Institutes Program (<http://clinicaltrials.gov/ct2/show/nCt00744315>). This is the first clinical gene therapy trial hosted in South America, in the midst of dozens of clinical trials promoted by multinational companies or foreign research institutions with the participation of researchers from South America (Table 7).

Table 7 – Sponsors of clinical gene therapy trials with participation of institutions in South America. Numbers in parentheses indicate the number of trials sponsored by the corresponding company

Actelion (5)
Bristol MyersSquibb (2)
Eli Lilly (1)
Genentech (3)
Glaxo Smith Kline (7)
Hoffman-La Roche (3)
Hoosier Oncology Group (1)
MedImmune LLC (1)
National Cancer Institute USA (1)
Office of Rare Diseases (1)
Sanofi-Aventis (9)
Shire Human Genetic Therapies Inc (1)
St Jude's Children's Research Hospital (2)
Cardiology Institute of RGS + Fapergs + CNPq-Instituto do Milênio Rede de Terapia Gênica (1) (Gene Therapy Network – Millennium Institute)

Source: NIH-USA (Clinicatrials.gov), Dec. 2009.

Gene therapy and biotechnology in Brazil

Consistent with the embryonic stage of gene therapy research, there is not much interest from the private sector in this area in the country. Recently, however, a company hosted in the high-technology Park Foundation was established in Petrópolis, in the state of Rio de Janeiro, which, among other services of biotechnology nature, is beginning to provide support for gene therapy trials in the country.

The vector for the clinical trial of myocardial revascularization started in Porto Alegre was produced by this service company, an unprecedented event in the country and a harbinger of new partnerships between the private sector and academic institutions in scientific research.

In turn, the awareness of the crucial role of mechanisms to protect intellectual property in this area is creating habits in researchers previously unconcerned with patenting products and processes of biotechnological interest. Thus, the performance of the Gene Therapy Network has also stimulated the first international filing of a gene therapy patent in its strict sense by a Brazilian institution, as a result of this author's empirical research (World Intellectual Property Organization - WO2009/121157 - PCt/BR2009/000093 .)

Conclusion

We are still in the early stages of gene therapy history and all that has been done to date are but the first steps on a long and winding road (Flotte, 2007). But now there are some occasional successes that demonstrate the feasibility of incorporating gene therapy into medical practice. The main advances, so far, are in the areas of hemophilia, certain types of cancer, severe combined immunodeficiency syndromes and certain retinopathies.

There has been great progress in the design and construction of new safer and more efficient vectors (Räty et al., 2008). In particular, the immune responses of patients are being studied in depth, new animal model studies have been developed and research is advancing towards increasing the safety of clinical trials.

The problems are not trivial. Just remember that, after all the medical progress made to date, despite the success achieved in new treatments and in the prevention of so many diseases in the last 150 years, we are still struggling against incurable diseases that challenge the imagination and scientific and technological competence of the entire scientific world.

There are, however, reasons for optimism, and the expectation of success in gene therapy technologies has gradually increased. A sign of the viability of gene therapy in the near future is the increasing investment of biotech companies in the development and filing of requests for the release of biological products relating to gene therapy.

Brazil is preparing to participate in the advent of gene therapy in clinical practice. The number of scientists, technicians, physicians and entrepreneurs

involved in this field in the country is still insignificant compared to First World countries. But from the financial and scientific and educational standpoints, the decision to invest in this area will certainly yield significant return to the Brazilian medicine of the twenty-first century.

References

AIUTI, A. et al. Gene therapy for immunodeficiency due to adenosine deaminase deficiency. *N. Engl. J. Med.*, v.360, n.5, p.447-58, 2009.

AIUTI, A.; RONCAROLO, M. G. Ten years of gene therapy for primary immune deficiencies. *Hematology Am. Soc. Hematol. Educ. Program*, p.682-9, 2009.

ANDERSON, W. F. Human gene therapy: the initial concepts. In: BRIGHAM, K. L. (Ed.) *Gene therapy for diseases of the lung*. S. l.: CRC Press, 1990. p. 3-16.

ANDERSON, W. F. et al. The ADA human gene therapy clinical protocol: Points to consider response with clinical protocol. *Hum. Gene Ther.*, v.1, n.3, p.331-62, 1990.

ANDREWES, C. Richard Edwin Shope. In: *Biographical memoirs*. s. l.: National Academy of Sciences, 1979. v.50, p.352-75.

ANDREWES, C. H. Rhinoviruses and common colds. *Annu. Rev. Med.*, v.17 p.361-70, 1966.

ATKINS, G. J. et al. Therapeutic and prophylactic applications of alphavirus vectors. *Expert Rev. Mol. Med.*, v.10, p.e33, 2008.

BAGLEY, J. et al. Gene therapy in type 1 diabetes. *Crit. Rev. Immunol.*, v.28, n.4, p.301-24, 2008.

BAINBRIDGE, J. W. et al. Effect of gene therapy on visual function in Leber's congenital amaurosis. *N. Engl. J. Med.*, v.358, n.21, p.2231-9, 2008.

BAUZON, M.; HERMISTON, T. W. Exploiting diversity: genetic approaches to creating highly potent and efficacious oncolytic viruses. *Curr. Opin. Mol. Ther.*, v.10, n.4, p.350-5, 2008.

BERETA, G. et al. Impact of retinal disease-associated RPE65 mutations on retinoid isomerization. *Biochemistry*, v.47, n.37, p.9856-65, 2008.

BLAESE, R. M. et al. T lymphocyte-directed gene therapy for ADA-SCID: initial trial results after 4 years. *Science*, v.270, n.5235, p.475-80, 1995.

BOBROW, M.; THOMAS, S. Patenting DNA. *Curr. Opin. Mol. Ther.*, v.4, n.6, p.542-7, 2002.

BRAAK, H.; DEL TREDICI, K. Assessing fetal nerve cell grafts in Parkinson's disease. *Nat. Med.*, v.14, n.5, p.483-5, 2008.

BRINKMAN, R. R. et al. Human monogenic disorders – a source of novel drug targets. *Nat. Rev. Genet.*, v.7, n.4, p.249-60, 2006.

- BUCKLEY, R. H. Molecular defects in human severe combined immunodeficiency and approaches to immune reconstitution. *Annu. Rev. Immunol.*, v.22, p.625-55, 2004.
- CACABELOS, R. Molecular pathology and pharmacogenomics in Alzheimer's disease: polygenic-related effects of multifactorial treatments on cognition, anxiety and depression. *Methods Find Exp. Clin. Pharmacol.*, v.29, Suppl A, p.1-91, 2007.
- CARDONE, M. Prospects for gene therapy in inherited neurodegenerative diseases. *Curr. Opin. Neurol.*, v.20, n.2, p.151-8, 2007.
- CATTANEO, R. et al. Reprogrammed viruses as cancer therapeutics: targeted, armed and shielded. *Nat. Rev. Microbiol.*, v.6, n.7, p.529-40, 2008.
- CAVA ZZANA-CALVO, M.; FISCHER, A. Gene therapy for severe combined immunodeficiency: are we there yet? *J. Clin. Invest.*, v.117, n.6, p.1456-65, 2007.
- CEMAZAR, M.; SERSA, G. Electrotransfer of therapeutic molecules into tissues. *Curr. Opin. Mol. Ther.*, v.9, n.6, p.554-62, 2007.
- CHEN, Q. et al. Gene therapy for Parkinson's disease: progress and challenges. *Curr. Gene Ther.*, v.5, p.71-80, 2005.
- CHRISTINE, C. W. et al.. Safety and tolerability of putaminal AADC gene therapy for Parkinson disease. *Neurology*, v.73, n.20, p.1662-9, 2009.
- CIDECIYAN, A. V. et al. Human gene therapy for RPE65 isomerase deficiency activates the retinoid cycle of vision but with slow rod kinetics. *Proc. Natl. Acad. Sci. USA*, v.105, n.39, p.15112-7, 2008.
- CLANCHY, F. I.; WILLIAMS, R. O. Plasmid DNA as a safe gene delivery vehicle for treatment of chronic inflammatory disease. *Expert Opin. Biol. Ther.*, v.8, n.10, p.1507-19, 2008.
- COLEMAN, W. B.; TSONGALIS, G. J. (Ed.) *Molecular pathology: the molecular basis of human disease*. S. l.: Academic Press, 2009. 664p.
- CULVER, K. W. et al. Correction of ADA deficiency in human T lymphocytes using retroviral-mediated gene transfer. *Transplant Proc.*, v.23, n.1 Pt. 1, p.170-1, 1991.
- DASS, C. R. Lipoplex-mediated delivery of nucleic acids: factors affecting in vivo transfection. *J. Mol. Med.*, v.82, n.9, p.579-91, 2004.
- DAUER, W.; PRZEDBORSKI, S. Parkinson's disease: mechanisms and models. *Neuron*, v.39, p.889-909, 2003.
- DAWSON, T. M.; DAWSON, V. L. Molecular pathways of neurodegeneration in Parkinson's disease. *Science*, v.302, p.819-22, 2003.
- DIAMOND, A.; JANKOVIC, J. The effect of deep brain stimulation on quality of life in movement disorders. *J. Neurol. Neurosurg Psychiatry*, v.76, p.1188-93, 2005.
- DORMOND, E. et al. From the first to the third generation adenoviral vector: what parameters are governing the production yield? *Biotechnol. Adv.*, v.27, n.2, p.133-44, 2009.

- EINSTEIN, O.; BEN-HUR, T. The changing face of neural stem cell therapy in neurologic diseases. *Arch. Neurol.*, v.65, n.4, p.452-6, 2008.
- EVAN, G.; LITTLEWOOD, T. A matter of life and cell death. *Science*, v.281, n.5381, p.1317-22, 1998.
- FAVA RD, C. et al. Electrotransfer as a non viral method of gene delivery. *Curr. Gene Ther.*, v.7, n.1, p.67-77, 2007.
- FISCHER, A.; CAVA ZZANA-CALVO, M Gene therapy of inherited diseases. *Lancet*, v.371, n.9629, p.2044-7, 2008.
- FJORD-LARSEN, L. et al. Efficient in vivo protection of nigral dopaminergic neurons by lentiviral gene transfer of a modified Neurturin construct. *Exp. Neurol.*, v.195, p.49-60, 2005.
- FLOTTE, T. R. Gene therapy: the first two decades and the current state-of-the-art. *J. Cell Physiol.*, v.213, n.2, p.301-5, 2007.
- FRIEDMANN T. The road toward human gene therapy--a 25-year perspective. *Ann Med.*, v.29 n.6 p.575-7, 1997.
- FRIEDMANN, T. Stanfield Rogers: insights into virus vectors and failure of an early gene therapy model. *Mol. Ther.*, v.4, n.4, p.285-8, 2001.
- GASPAR, H. B. et al. Gene therapy of X-linked severe combined immunodeficiency by use of a pseudotyped gammaretroviral vector. *Lancet*, v.364, n.9452, p.2181-7, 2004.
- GAUTHIER, S.; POIRIER, J. Current and future management of Alzheimer's disease. *Alzheimers Dement.*, v.4, n.1, Suppl. 1, p.S48-50, 2008.
- GILL, D. R. et al. Progress and prospects: the design and production of plasmid vectors. *Gene Ther.*, v.16, n.2, p.165-71, 2009.
- GOLDSTEIN, M. Decade of the brain: an agenda for the nineties. *West. J. Med.*, v.161, p.239-41, 1994.
- GREEN, D. R.; EVAN, G. I. A matter of life and death. *Cancer Cell.*, v.1, n.1, p.19-30, 2002.
- GRIBBEN, J. G. Stem cell transplantation in chronic lymphocytic leukemia. *Biol. Blood Marrow Transplant.*, v.15, n.1, Suppl., p.53-8, 2008.
- GUTTMAN, M. et al. Current concepts in the diagnosis and management of Parkinson's disease. *CMAJ*, v.168, p.293-301, 2003.
- HACEIN-BEY-ABINA, S. et al. Gene therapy of X-linked severe combined immunodeficiency. *Int. J. Hematol.*, v.76, n.4, p.295-8, 2002.
- HACEIN-BEY-ABINA, S. et al. LMO2-associated clonal T cell proliferation in two patients after gene therapy for SCID-X1. *Science*, v.302, n.5644, p.415-9, 2003.

- HAN, J. J.; MCDONALD, C. M. Diagnosis and clinical management of spinal muscular atrophy. *Phys. Med. Rehabil. Clin. N. Am.*, v.19, n.3, p.661-80, 2008.
- HASBROUCK, N. C.; HIGH, K. A. AAV -mediated gene transfer for the treatment of hemophilia B: problems and prospects. *Gene Ther.*, v.15, n.11, p.870-5, 2008.
- HAUSER, O. et al. Encapsulated, genetically modified cells producing in vivo therapeutics. *Curr. Opin. Mol. Ther.*, v.6, n.4, p.412-20, 2004.
- HAUSWIRTH, W. et al. Treatment of Leber congenital amaurosis due to RPE65 mutations by ocular subretinal injection of adeno-associated virus gene vector: short-term results of a phase I trial. *Hum. Gene Ther.*, v.19, n.10, p.979-90, 2008.
- HERWEIJER, H.; WOLFF, J. A. Gene therapy progress and prospects: hydrodynamic gene delivery. *Gene Ther.*, v.14, n.2, p.99-107, 2007.
- HOLLANDER, A. I. den et al. FP Leber congenital amaurosis: genes, proteins and disease mechanisms. *Prog. Retin Eye Res.*, v.27, n.4, p.391-419, 2008.
- HOWE, S. J. et al. Insertional mutagenesis combined with acquired somatic mutations causes leukemogenesis following gene therapy of SCID-X1 patients. *J. Clin. Invest.*, v.118, n.9, p.3143-50, 2008.
- IMMONEN, A. et al. AdvHSV-tk gene therapy with intravenous ganciclovir improves survival in human malignant glioma: a randomised, controlled study. *Mol. Ther.*, v.10, n.5, p.967-72, 2004.
- JACKSON D. A.; SYMONS R. H., BERG, P. Biochemical method for inserting new genetic information into DNA of Simian Virus 40: circular SV40 DNA molecules containing lambda phage genes and the galactose operon of Escherichia coli. *Proc. Natl. Acad. Sci. USA.* v.69 n.10 p.2904-9, 1972.
- JALBERT, J. J. et al. Dementia of the Alzheimer type. *Epidemiol. Rev.*, v.30, p.15-34, 2008.
- KANG, U. J. et al. Gene therapy for Parkinson's disease: determining the genes necessary for optimal dopamine replacement in rat models. *Hum. Cell.*, v.14, n.1, p.39-48, 2001.
- KAPLITT, M. G. et al. Safety and tolerability of gene therapy with an adeno-associated virus (AAV) borne GAD gene for Parkinson's disease: an open label, phase I trial. *Lancet*, v.369, n.9579, p.2097-105, 2007.
- KOHN, D. B.; CANDOTTI, F. Gene therapy fulfilling its promise. *N. Engl. J. Med.*, v.360, n.5, p.518-21, 2009.
- KORDOWER, J. H. et al. Lewy body-like pathology in long-term embryonic nigral transplants in Parkinson's disease. *Nat. Med.*, v.14, n.5, p.504-6, 2008.
- LANDER, E. S. et al. Initial sequencing and analysis of the human genome. *Nature*, v.409, n.6822, p.860-921, 2001.
- LI, J. Y. et al. Lewy bodies in grafted neurons in subjects with Parkinson's disease suggest host-to-graft disease propagation. *Nat. Med.*, v.14, n.5, p.501-3, 2008.

- LINDEN, R. *Genes contra doenças*. Terapia gênica: uma nova era na genética. Rio de Janeiro: Vieira e Lent, 2008. 128p.
- LINDEN, R.; LENZ, G. Terapia gênica em neurologia. In: MORALES, M. M. (Ed.) *Terapias avançadas*. Rio de Janeiro: Atheneu, 2007. p.205-22.
- LINDVALL, O.; WAHLBERG, L. U. Encapsulated cell biodelivery of GDNF: a novel clinical strategy for neuroprotection and neuroregeneration in Parkinson's disease? *Exp. Neurol.*, v.209, n.1, p.82-8, 2008.
- LUNDBERG, C. et al. Applications of lentiviral vectors for biology and gene therapy of neurological disorders. *Curr. Gene Ther.*, v.8, n.6, p.461-73, 2008.
- LUO J. et al. Subthalamic GAD gene therapy in a Parkinson's disease rat model. *Science*, v.298, p.425-9, 2002.
- MACHIDA, C. A. (Ed.) *Viral vectors for gene therapy: methods and protocols*. S. 1.: Humana Press, 2002. 608p.
- MAGUIRE, A. M. et al. Safety and efficacy of gene transfer for Leber's congenital amaurosis. *N. Engl. J. Med.*, v.358, n.21, p.2240-8, 2008.
- MANNO, C. S. et al. AAV -mediated factor IX gene transfer to skeletal muscle in patients with severe hemophilia B. *Blood*, v.101, n.8, p.2963-72, 2003.
- MANNO, C. S. et al. Successful transduction of liver in hemophilia by AAV -Factor IX and limitations imposed by the host immune response. *Nat. Med.*, v.12, n.3, p.342-7, 2006.
- MARKS, W. J. et al. Safety and tolerability of intraputaminial delivery of CERE-120 (adeno-associated virus serotype 2-neurturin) to patients with idiopathic Parkinson's disease: an open-label, phase I trial. *Lancet Neurol.*, v.7, n.5, p.400-8, 2008.
- MENDEZ, I. et al. Dopamine neurons implanted into people with Parkinson's disease survive without pathology for 14 years. *Nat. Med.*, v.14, n.5, p.507-9, 2008.
- MERCOLA, K. E.; CLINE, M. J. Sounding boards. The potentials of inserting new genetic information. *N. Engl. J. Med.*, v.303, n.22, p.1297-300, 1980.
- MINGOZZI, F.; HIGH, K. A. Immune responses to AAV in clinical trials. *Curr. Gene Ther.*, v.7, n.5, p.316-24, 2007.
- MULLEN C. A. et al. Molecular analysis of T lymphocyte-directed gene therapy for adenosine deaminase deficiency: long-term expression in vivo of genes introduced with a retroviral vector. *Hum. Gene Ther.*, v.7 n.9 p.1123-9, 1996.
- MUUL, L. M. et al. Persistence and expression of the adenosine deaminase gene for 12 years and immune reaction to gene transfer components: long-term results of the first clinical gene therapy trial. *Blood*, v.101, n.7, p.2563-9, 2003.
- NAKANO, K. Neural circuits and topographic organization of the basal ganglia and related regions. *Brain Dev.*, v.22, Suppl. 1, p.S5-16, 2000.

NAT HWANI, A. C. et al. A review of gene therapy for haematological disorders. *Br. J. Haematol.*, v.28, n.1, p.3-17, 2005.

NCBI. Genes and disease. 2009. Available at: <<http://www.ncbi.nlm.nih.gov/books/bv.fcgi?rid=gnd&ref=sidebar>>.

O'CONNOR, T. P.; CRYSTA L, R. G. Genetic medicines: treatment strategies for hereditary disorders. *Nat. Rev. Genet.*, v.7, n.4, p.261-76, 2006.

OLANOW, C. W. et al. Why have we failed to achieve neuroprotection in Parkinson's disease? *Ann Neurol.*, v.64 Suppl 2, p.S101-10, 2008.

PARDRIDGE, W. M. Tyrosine hydroxylase replacement in experimental Parkinson's disease with transvascular gene therapy. *NeuroRx.*, v.2, n.1, p.129-38, 2005.

PARDRIDGE, W. M. Blood-brain barrier delivery of protein and non-viral gene therapeutics with molecular Trojan horses. *J. Control. Release*, v.122, n.3, p.345-8, 2007.

PEARSON, S. et al. China approves first gene therapy. *Nature Biotechnol.*, v.22, p.3-4, 2004.

PHACILITATE , Inc. 5th Annual Cell and Gene Therapy Forum. Washington, DC, 26-28 January, 2009.

POEHNER, W. J. et al. A homozygous deletion in RPE65 in a small Sardinian family with autosomal recessive retinal dystrophy. *Mol. Vis.*, v.6, p.192-8, 2000.

POEWE, W. Treatments for Parkinson disease - past achievements and current clinical needs. *Neurology*, v.72, n.7, Supp. p.S65-73, 2009.

PORTEUS, M. H. et al. A look to future directions in gene therapy research for monogenic diseases. *PLoS Genet.*, v.2, n.9, p.e133, 2006.

PROCHOWNIK, E. V. c-Myc: linking transformation and genomic instability. *Curr. Mol. Med.*, v.8, n.6, p.446-58, 2008.

RADUNOVIC, A. et al. Clinical care of patients with amyotrophic lateral sclerosis. *Lancet Neurol.*, v.6, n.10, p.913-25, 2007.

RAINOV, N. G.; REN, H. Gene therapy for human malignant brain tumors. *Cancer J.*, v.9, n.3, p.180-8, 2003.

RAPER, S. E. et al. Fatal systemic inflammatory response syndrome in an ornithine transcarbamylase deficient patient following adenoviral gene transfer. *Mol. Genet. Metab.*, v.80, n.1-2, p.148-58, 2003.

RAPER, S. E. et al. Developing adenoviral-mediated in vivo gene therapy for ornithine transcarbamylase deficiency. *J. Inherit Metab. Dis.*, v.21, Suppl. 1, p.119-37, 1998.

RÄTY, J. K. et al. Improving safety of gene therapy. *Curr. Drug Saf.*, v.3, n.1, p.46-53, 2008.

REFFELMANN, T. et al. Promise of blood- and bone marrow-derived stem cell transplantation for functional cardiac repair: putting it in perspective with existing therapy. *J. Am. Coll. Cardiol.*, v.53, n.4, p.305-8, 2008.

RIBACKA, C. et al. Cancer, stem cells, and oncolytic viruses. *Ann. Med.*, v.40, n.7, p.496-505, 2008.

ROGERS, S.; ROUS, P. Joint action of a chemical carcinogen and a neoplastic virus to induce cancer in rabbits; results of exposing epidermal cells to a carcinogenic hydrocarbon at time of infection with the Shope papilloma virus. *J. Exp. Med.*, v.93, n.5, p.459-88, 1951.

ROGERS, S. Serial transplantation of rabbit papillomas caused by the Shope virus. *J. Exp. Med.*, v.95, n.6, p.543-54, 1952.

SANVICENS, N.; MARCO, M. P. Multifunctional nanoparticles--properties and prospects for their use in human medicine. *Trends Biotechnol.*, v.26, n.8, p.425-33, 2008.

SCAGLIA, F.; LEE, B. Clinical, biochemical, and molecular spectrum of hyperargininemia due to arginase I deficiency. *Am. J. Med. Genet. C. Semin. Med. Genet.*, v.142C, n.2, p.113-20, 2006.

SILVA, C. L. et al. Recent advances in DNA vaccines for autoimmune diseases. *Expert Rev. Vaccines*, v.8, n.2, p.239-52, 2009.

SYKES, K. Progress in the development of genetic immunization. *Expert Rev. Vaccines*, v.7, n.9, p.1395-404, 2008.

THAJEB, P. et al. The effects of storage conditions and trophic supplementation on the survival of fetal mesencephalic cells. *Cell Transplant.*, v.6, p.297-307, 1997.

TORRENTE, Y.; POLLI, E. Mesenchymal stem cell transplantation for neurodegenerative diseases. *Cell Transplant.*, v.17, n.10-11, p.1103-13, 2008.

Van der SPUIJ, J. et al. Predominant rod photoreceptor degeneration in Leber congenital amaurosis. *Mol Vis.*, v.11, p.542-53, 2005.

VA ZQUEZ, A. et al. The genetics of the p53 pathway, apoptosis and cancer therapy. *Nat. Rev. Drug Discov.*, v.7, n.12, p.979-87, 2008.

VELLAI, T.; VIDA, G. The origin of eukaryotes: the difference between prokaryotic and eukaryotic cells. *Proc. Biol. Sci.*, v.266, n.1428, p.1571-7, 1999.

VENTER, J. C. et al. The sequence of the human genome. *Science*, v.291, n.5507, p.1304-51, 2001.

VOSS, C. Production of plasmid DNA for pharmaceutical use. *Biotechnol. Annu. Rev.*, v.13, p.201-22, 2007.

WANG, X. et al. Mitotic checkpoint defects in human cancers and their implications to chemotherapy. *Front Biosci.*, v.13, p.2103-14, 2008.

WATSON , J. D. et al. *Recombinant DNA: genes and genomics: a short course*. S. l.: Freeman, 2006. 474p.

WU, C.; LU, Y. Inclusion of high molecular weight dextran in calcium phosphate mediated transfection significantly improves gene transfer efficiency. *Cell. Mol. Biol.*, Noisy-le-grand, v.53, n.4, p.67-74, 2007.

YI, Y.; HAHM, S. H.; LEE, K. H. Retroviral gene therapy: safety issues and possible solutions. *Curr. Gene Ther.*, v.5 n.1 p.25-35, 2005.

ABSTRACT – Gene therapy is the therapeutic procedure based on the introduction of healthy genes using recombinant DNA techniques. The first successful clinical trial of this technique was published in 1990. Despite the occurrence of adverse effects in certain clinical trials, some of them serious, both laboratories and companies are continuously developing novel materials and establishing both safer and more effective procedures. Although still in experimental stages, recent progress points to growing opportunities for investment by industry, as well as justify the expectation that, in some cases, this technology may reach clinical practice within a few years.

KEYWORDS: Genes, Genetic diseases, Genetic engineering, Genome manipulation, Advanced therapies, Biotechnology.

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